# ITGAL and PSMC5 are promising druggable targets for treating Colorectal Neoplasms that control activity of FOSL1, ZEB1 and KLF8 transcription factor on promoters of genes carrying sequence variations

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Genome Enhancer release 3.1 (TRANSFAC®, TRANSPATH® and HumanPSD™ release 2022.2)









### **Abstract**

In the present study we applied the software package "Genome Enhancer" to a data set that contains *genomics* data. The study is done in the context of *Colorectal Neoplasms*. The goal of this pipeline is to identify potential drug targets in the molecular network that governs the studied pathological process. In the first step of analysis pipeline discovers transcription factors (TFs) that regulate genes activities in the pathological state. The activities of these TFs are controlled by so-called master regulators, which are identified in the second step of analysis. After a subsequent druggability checkup, the most promising master regulators are chosen as potential drug targets for the analyzed pathology. At the end the pipeline comes up with (a) a list of known drugs and (b) investigational active chemical compounds with the potential to interact with selected drug targets.

From the data set analyzed in this study, we found the following TFs to be potentially involved in the regulation of the genes carrying sequence variations: FOSL1, ZEB1 and KLF8. The subsequent network analysis suggested

- · integrins
- PML
- · 26S proteasome

as the most promising molecular targets for further research, drug development and drug repurposing initiatives on the basis of identified molecular mechanism of the studied pathology. Having checked the actual druggability potential of the full list of identified targets, both, via information available in medical literature and via cheminformatics analysis of drug compounds, we have identified the following drugs as the most promising treatment candidates for the studied pathology: Erlotinib, seliciclib, Camptothecin and 2,6-Dihydroanthra/1,9-Cd/Pyrazol-6-One.

### 1. Introduction

Recording "-omics" data to measure gene activities, protein expression or metabolic events is becoming a standard approach to characterize the pathological state of an affected organism or tissue. Increasingly, several of these methods are applied in a combined approach leading to large "multiomics" datasets. Still the challenge remains how to reveal the underlying molecular mechanisms that render a given pathological state different from the norm. The disease-causing mechanism can be described by a re-wiring of the cellular regulatory network, for instance as a result of a genetic or epigenetic alterations influencing the activity of relevant genes. Reconstruction of the disease-specific regulatory networks can help identify potential master regulators of the respective pathological process. Knowledge about these master regulators can point to ways how to block a pathological regulatory cascade. Suppression of certain molecular targets as components of these cascades may stop the pathological process and cure the disease.

Conventional approaches of statistical "-omics" data analysis provide only very limited information about the causes of the observed phenomena and therefore contribute little to the understanding of the pathological molecular mechanism. In contrast, the "upstream analysis" method [1-4] applied here has been deviced to provide a casual interpretation of the data obtained for a pathology state. This approach comprises two major steps: (1) analysing promoters and enhancers of genes carrying sequence variations for the transcription factors (TFs) involved in their regulation and, thus, important for the process under study; (2) re-constructing the signaling pathways that activate these TFs and identifying master regulators at the top of such pathways. For the first step, the database TRANSFAC® [6] is employed together with the TF binding site identification algorithms Match [7] and CMA [8]. The second step involves the signal transduction database TRANSPATH® [9] and special graph search algorithms [10] implemented in the software "Genome Enhancer".

The "upstream analysis" approach has now been extended by a third step that reveals known drugs suitable to inhibit (or activate) the identified molecular targets in the context of the disease under study. This step is performed by using information from

HumanPSD $^{\text{TM}}$  database [5]. In addition, some known drugs and investigational active chemical compounds are subsequently predicted as potential ligands for the revealed molecular targets. They are predicted using a pre-computed database of spectra of biological activities of chemical compounds of a library of 2245 known drugs and investigational chemical compounds from HumanPSD $^{\text{TM}}$  database. The spectra of biological activities for these compounds are computed using the program PASS on the basis of a (Q)SAR approach [11-13]. These predictions can be used for the research purposes - for further drug development and drug repurposing initiatives.

### 2. Data

For this study the following experimental data was used:

Table 1. Experimental datasets used in the study

File name	Data type
CRC_variants	Genomics

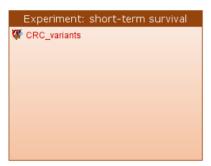


Figure 1. Annotation diagram of experimental data used in this study. With the colored boxes we show those sub-categories of the data that are compared in our analysis.

### 3. Results

We have analyzed the following condition: Experiment: short-term survival.

### 3.1. Identification of target genes

In the first step of the analysis *target genes* were identified from the uploaded experimental data. The most frequently mutated genes were used as target genes.

Table 2. Top ten the most frequently mutated genes in Experiment: short-term survival.

See full table →						
ID	Gene description	Gene symbol	Gene schematic representation	Number of variations	Gene weight	Weighted score
ENSG00000132570	pterin-4 alpha-carbinolamine dehydratase 2	PCBD2		172	419.42	629.12
ENSG00000228716	dihydrofolate reductase	DHFR		56	131.75	395.25
ENSG00000067057	phosphofructokinase, platelet	PFKP	***************************************	92	223.43	335.15
ENSG0000003402	CASP8 and FADD like apoptosis regulator	CFLAR	-mina-indichtria dial-	43	111.41	334.23
ENSG00000176890	thymidylate synthetase	TYMS		44	110.62	331.86
ENSG00000169894	mucin 3A, cell surface associated	MUC3A		68	162.02	324.05
ENSG00000242086	MUC20 overlapping transcript	MUC20- OT1	+	147	323.51	323.51
ENSG00000204525	major histocompatibility complex, class I, C	HLA-C		71	154.6	309.21
ENSG00000248923	MT-ND5 pseudogene 11	MTND5P11		126	306.95	306.95
ENSG00000134086	von Hippel-Lindau tumor suppressor	VHL		43	98.24	294.71

### 3.2. Functional classification of genes

A functional analysis of genes carrying sequence variations was done by mapping the genes to several known ontologies, such as Gene Ontology (GO), disease ontology (based on HumanPSD™ database) and the ontology of signal transduction and metabolic pathways from the TRANSPATH® database. Statistical significance was computed using a binomial test. Figures 2-4 show the most significant categories.

# The most frequently mutated genes in Experiment: short-term survival:

300 top mutated genes were taken for the mapping.

# GO (biological process)

biological\_process Gene Ontology treemap

						<u> </u>		
positive regulation of protein metabolic process	regulation of protein metabolic process	regulation of protein modification process	regulation of signal transduction	regulation of signaling	positive regulation of cellular process	circulatory system development	response to chemical	cell surface receptor signaling pathway
					positive regulat of cellular proce	Circulatory syste	em response to chemica	cell surface receptor
positive regulation of cellular protein metabolic process	positive regulation of protein modification	regulation of cellular protein metabolic process	regulation of sign		cellular metabolic process	response to stress	organic cyclic compound metabolic process	anatomical structure morphogenesis
regulation of p	process rotein metabo	lic process	cytokine-mediated signaling pathway	signaling pathway	cellular metabolic process	response to stress	organic cyclic compound metabolic process	anatomical structure morphogenesis
positive regulation of metabolic proce	11	julation of nitrogen metabolic process			regulation of cellular component organization	organonitrogen compound metabolic process	regulation of metabolic process	regulation of primary metabolic process
						organonitrogen compound metabolic process	regulation of metabolic process	regulation of primary metabolic process
positive regulation	01	ve regulation	type I interferon sig	signaling pathway	regulation of cellular component organization	metabolic process	cardiovascular system development	negative regulation of biological process
cellular metabolic pro	0000	cromolecule polic process	positive regulation of	biological process	regulation of response to stimulus	metabolic process	cardiovascular system development	negative regulation of biological process
positive regula	tion of metabo	lic process			regulation of	regulation of nitrogen compound metabolic process regulation of	system development	primary metabolic process
response to organic sub		ar response to nic substance	positive regulation of cellular response to		response to stimulus  cell population  proliferation	nitrogen compound metabolic process	system development	primary metabolic process
						vasculature development	regulation of cellular metabolic process	nitrogen compound metabolic process
					cell population		regulation of cellular	nitrogen compound
response to	organic su	ubstance	cellular response to	chemical stimulus	proliferation	vasculature development	metabolic process	metabolic process

Figure 2. Enriched GO (biological process) of the most frequently mutated genes in Experiment: short-term survival. Full classification  $\rightarrow$ 

# TRANSPATH® Pathways (2022.2)

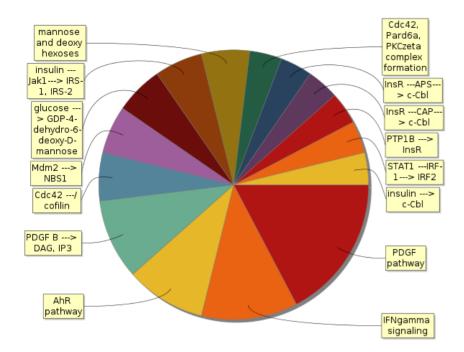


Figure 3. Enriched TRANSPATH® Pathways (2022.2) of the most frequently mutated genes in Experiment: short-term survival. Full classification  $\rightarrow$ 

### HumanPSD(TM) disease (2022.2)

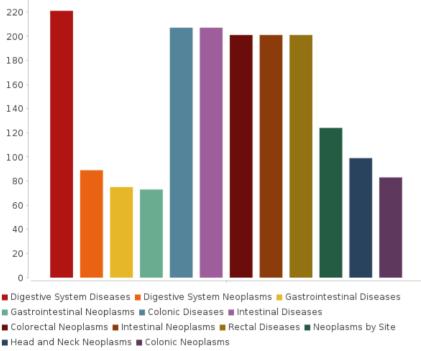
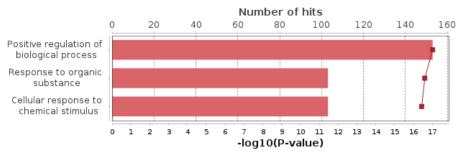


Figure 4. Enriched HumanPSD(TM) disease (2022.2) of the most frequently mutated genes in Experiment: short-term survival. The size of the bars correspond to the number of biomarkers of the given disease found among the input set.

Full classification  $\rightarrow$ 

The result of overall Gene Ontology (GO) analysis of the genes carrying sequence variations of the studied pathology can be summarized by the following diagram, revealing the most significant functional categories overrepresented among the observed (genes carrying sequence variations):



- The most frequently mutated genes in Experiment: short-term survival hits
- The most frequently mutated genes in Experiment: short-term survival -log10(P-value)

### 3.3. Analysis of enriched transcription factor binding sites and composite modules

In the next step a search for transcription factors binding sites (TFBS) was performed in the regulatory regions of the *target genes* by using the TF binding motif library of the TRANSFAC® database. We searched for so called **composite modules** that act as potential condition-specific **enhancers** of the *target genes* in their upstream regulatory regions (-1000 bp upstream of transcription start site (TSS)) and identify transcription factors regulating activity of the genes through such **enhancers**.

Classically, **enhancers** are defined as regions in the genome that increase transcription of one or several genes when inserted in either orientation at various distances upstream or downstream of the gene [8]. Enhancers typically have a length of several hundreds of nucleotides and are bound by multiple transcription factors in a cooperative manner [9].

In the current work, we use the Genomics data from the "Yes VCF track" track to predict positions of potential *enhancers* where the observed sequence variations may influence the gene expression in the pathology under study. We scan 5kb flanking regions and the body of all genes caring the variations, with a sliding window of 1100bp size and find the position of the window with the maximal sum of the mutation weights, where we then perform the search for potential condition-specific enhancers (CMA model search).

We analyzed mutations that were revealed in the potential enhancers located upstream, downstream or inside the *target genes* (see Table 3). We identified 23644 mutations potentially affecting gene regulation. Table 4 shows the following lists of PWMs whose sites were lost or gained due to these mutations. Weighting of mutations was done in respect to the significance of the change in TF affinity binding to the sequence. Mutations that maximally affected the change of binding affinity received higher weights. These PWMs were put in focus of the CMA algorithm that constructs the model of the enhancers by specifying combinations of TF motifs (see more details of the algorithm in the Methods section).

Table 3. Mutations revealed in the most frequently mutated genes

See full table $\rightarrow$			
ID	Gene symbol	<b>Gene schematic representation</b>	Number of variations
ENSG00000132570	PCBD2		660
ENSG00000248923	MTND5P11		459
ENSG00000247627	MTND4P12	# # # # # # # # # # # # # # # # # # #	374
ENSG00000230021	ENSG00000230021	**************************************	360
ENSG00000249119	MTND6P4		279
ENSG00000242086	MUC20-OT1	+1111-4141-161-16141-1614-11-1611-1911-19	252
ENSG00000198868	MTND4LP30	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	245
ENSG00000263963	ENSG00000263963	11 11 1 <b>111</b> 111 111 11 11 11 11 11 11 11 11 11	245
ENSG00000154237	LRRK1	111111111111111111111111111111111111111	230
ENSG00000244921	MTCYBP18	1111	225

Table 4. PWMs whose sites were lost or gained due to mutations in the most frequently mutated genes **See full table**  $\rightarrow$ 

ID	P-value (gains)	P-value (losses)	yesCount (gains)	yesCount (losses)
V\$OTX2_06	3.65E-2	8.72E-14	25	70
V\$GSC_04	3.12E-2	3.07E-11	5	82
V\$ZBTB33_07	1.06E-2	8.27E-12	13	611
V\$ZBTB33_05	6.47E-3	6.92E-10	12	535
V\$MATH1_Q2	1.99E-3	3.66E-12	204	389
V\$E2A_03	1.6E-3	1.14E-12	231	387
V\$MAFG_12	3.44E-5	3.47E-10	34	1295
V\$E2F8_03	1.22E-6	1.32E-9	63	1618
V\$MYBL2_04	1.81E-12	1.1E-10	2161	2211
V\$E2F1DP1RB_01	1.88E-25	1.47E-9	3568	3831
V\$E2F4DP1_01	1.88E-25	1.14E-10	3568	4126
V\$SP1_02	1.18E-26	1.56E-2	250	6
V\$KLF8_Q5	6.06E-27	6.79E-3	313	214
V\$PAX5_Q6	4.25E-27	1.83E-3	1935	2567
V\$E2F1_Q6	3.25E-28	3.18E-2	2073	14
V\$E2F1DP2_01	2.9E-31	1.81E-11	3552	3615
V\$E2F3_12	1.1E-31	3.12E-2	1493	5
V\$E2F_Q4_02	1.46E-35	2.93E-2	2015	55
V\$E2F1_04	2.31E-36		903	
V\$E2F3_09	1.41E-41		3966	

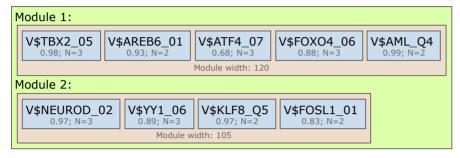
We applied the Composite Module Analyst (CMA) [8] method to detect such potential enhancers, as targets of multiple TFs bound in a cooperative manner to the regulatory regions of the genes of interest. CMA applies a genetic algorithm to construct a generalized model of the enhancers by specifying combinations of TF motifs (from TRANSFAC®) whose sites are most frequently clustered together in the regulatory regions of the studied genes. CMA identifies the transcription factors that through their cooperation provide a synergistic effect and thus have a great influence on the gene regulation process.

# Enhancer model potentially involved in regulation of target genes (the most frequently mutated genes in Experiment: short-term survival).

To build the most specific composite modules we choose top mutated genes as the input of CMA algorithm. The obtained CMA model is then applied to compute CMA score for all the most frequently mutated genes in Experiment: short-term survival.

The model consists of 2 module(s). Below, for each module the following information is shown:

- PWMs producing matches,
- number of individual matches for each PWM,
- score of the best match.



Model score (-p\*log10(pval)): 25.30 Wilcoxon p-value (pval): 1.24e-49

Penalty (p): 0.517

Average yes-set score: 8.33 Average no-set score: 6.50

**AUC:** 0.77

Separation point: 7.38 False-positive: 30.69% False-negative: 26.33%

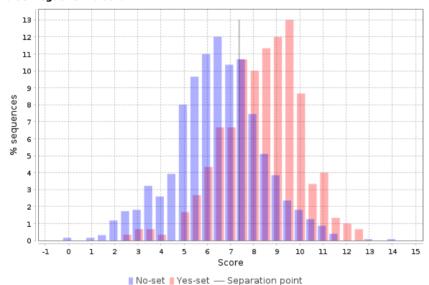


Table 5. List of top ten the most frequently mutated genes in Experiment: short-term survival with identified enhancers in their regulatory regions. **CMA score** - the score of the CMA model of the enhancer identified in the regulatory region. **See full table**  $\rightarrow$ 

Ensembl IDs	Gene symbol	Gene description	CMA score	Factor names
ENSG00000179477	ALOX12B	arachidonate 12-lipoxygenase, 12R type	14.95	ATF-4(h), TBX2(h), Runx1(h),Runx2(h),Runx3(h), ZEB1(h), Fra-1(h), foxo4(h), YY1(h)
ENSG00000214999		novel transcript, antisense to ALOX12B	14.95	ATF-4(h), TBX2(h), Runx1(h),Runx2(h),Runx3(h), ZEB1(h), Fra-1(h), foxo4(h), YY1(h)
ENSG00000182858	ALG12	ALG12 alpha-1,6- mannosyltransferase	14.75	KLF8(h), YY1(h), Fra-1(h), ZEB1(h), ATF-4(h), foxo4(h), Runx1(h),Runx2(h),Runx3(h)
ENSG00000273192		novel transcript, antisense to ALG12	14.55	Fra-1(h), ZEB1(h), ATF-4(h), foxo4(h), Runx1(h),Runx2(h),Runx3(h), TBX2(h), YY1(h)
ENSG00000265558	MIR3918	microRNA 3918	14.31	YY1(h), Fra-1(h), ATF-4(h), TBX2(h), foxo4(h), ZEB1(h), Runx1(h),Runx2(h),Runx3(h)
ENSG00000111110	PPM1H	protein phosphatase, Mg2+/Mn2+ dependent 1H	14.05	Runx1(h),Runx2(h),Runx3(h), ATF-4(h), foxo4(h), TBX2(h), ZEB1(h), KLF8(h), YY1(h)
ENSG00000224884		novel transcript	13.9	foxo4(h), ZEB1(h), TBX2(h), ATF-4(h), Runx1(h),Runx2(h),Runx3(h), KLF8(h), Fra-1(h)
ENSG00000114656	CFAP92	cilia and flagella associated protein 92 (putative)	13.9	NeuroD1(h), YY1(h), Fra-1(h), KLF8(h), TBX2(h), foxo4(h), ZEB1(h)
ENSG00000102158	MAGT1	magnesium transporter 1	13.84	YY1(h), Fra-1(h), KLF8(h), ZEB1(h), foxo4(h), TBX2(h), Runx1(h),Runx2(h),Runx3(h)
ENSG00000179776	CDH5	cadherin 5	13.79	YY1(h), Fra-1(h), Runx1(h),Runx2(h),Runx3(h), TBX2(h), ZEB1(h), foxo4(h), ATF-4(h)

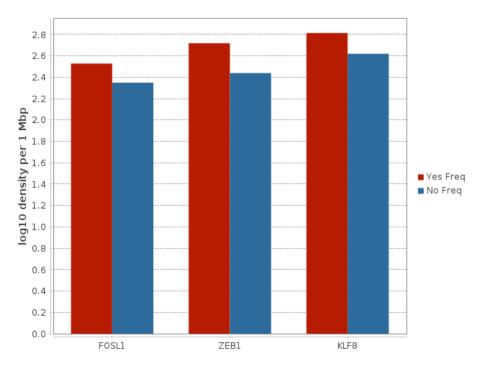
On the basis of the enhancer models we identified transcription factors potentially regulating the **target genes** of our interest. We found 11 transcription factors controlling expression of the genes associated with genomic variations (see Table 6).

Table 6. Transcription factors of the predicted enhancer model potentially regulating the genes carrying sequence variations (the most frequently mutated genes in Experiment: short-term survival). **Yes-No ratio** is the ratio between frequencies of the sites in Yes sequences versus No sequences. It describes the level of the enrichment of binding sites for the indicated TF in the regulatory target regions. **Regulatory score** is the measure of involvement of the given TF in the controlling of expression of genes that encode master regulators presented below (through positive feedback loops).

### See full table $\rightarrow$

ID	Gene symbol	Gene description	Regulatory score	Yes-No ratio
MO000025684	FOSL1	FOS like 1, AP-1 transcription factor subunit	6.92	1.51
MO000139677	ZEB1	zinc finger E-box binding homeobox 1	6.26	1.9
MO000095459	KLF8	Kruppel like factor 8	6.26	1.56
MO000078913	YY1	YY1 transcription factor	6.06	1.24
MO00000904	FOXO4	forkhead box O4	5.78	2.81
MO000028384	NEUROD1	neuronal differentiation 1	5.28	1.45
MO000025375	RUNX1	RUNX family transcription factor 1	5.27	1.86
MO000019140	ATF4	activating transcription factor 4	5.08	2.14
MO000026238	RUNX3	RUNX family transcription factor 3	5.06	9.19
MO000026285	RUNX2	RUNX family transcription factor 2	4.5	2.26

The following diagram represents the key transcription factors, which were predicted to be potentially regulating genes carrying sequence variations in the analyzed pathology: FOSL1, ZEB1 and KLF8.



# 3.4. Finding master regulators in networks

In the second step of the upstream analysis common regulators of the revealed TFs were identified. We identified 174 signaling proteins whose structure and function is highly damaged by the mutations (see Table 7).

Table 7. Signaling proteins whose structure and function are damaged by the mutations in the most frequently mutated genes

See full table  $\rightarrow$ 

ID	Title	<b>Mutation count</b>	Consequence	Codons
MO000138949	Drp1(h)	13	NMD_transcript_variant,stop_gained	Gaa/Taa
MO000206935	C11orf74(h)	12	stop_gained	Gaa/Taa
MO000211774	DPAGT1(h)	10	NMD_transcript_variant,frameshift_variant	ttc/ttTc
MO000019673	p85alpha(h)	9	stop_gained	Cga/Tga
MO000190658	GPSM2(h)	9	stop_gained	Gaa/Taa
MO000093071	chd8(h)	8	stop_gained	taC/taA
MO000113258	MYPT1(h)	8	NMD_transcript_variant,frameshift_variant	aga/aAga
MO000127741	SMC4L1(h)	8	stop_gained	Cga/Tga
MO000214698	MS4A6A(h)	8	NMD_transcript_variant,frameshift_variant	-/T,tta/ttTa
MO000035319	kinectin(h)	7	NMD_transcript_variant,frameshift_variant	-/A

Top 100 mutated proteins for the most frequently mutated genes were used in the algorithm of master regulator search as a list of nodes of the signal transduction network that are removed from the network during the search of master regulators (see more details about the algorithm in the Methods section). These master regulators appear to be the key candidates for therapeutic targets as they have a master effect on regulation of intracellular pathways that activate the pathological process of our study. The identified master regulators are shown in Table 8.

Table 8. Master regulators that may govern the regulation of the most frequently mutated genes in Experiment: short-term survival. **Total rank** is the sum of the ranks of the master molecules sorted by keynode score, CMA score, genomics data.

See full table →

occ run tubic	,				
ID	Master molecule name	Gene symbol	Gene description	Total rank	Weighted score
MO000079043	PML-4(h)	PML	PML nuclear body scaffold	177	136.46
MO000017291	integrins	ITGA1, ITGA2B, ITGA3, ITGA4, ITGA5, ITGA6, ITGA8, ITGA9, ITGAL, ITGAV, ITGB1, ITGB2, ITGB3, ITGB4, I	integrin subunit alpha 1, integrin subunit alpha 2b, integrin subunit alpha 3, integrin subunit alph	255	191.78
MO000045014	PML(h)	PML	PML nuclear body scaffold	301	136.46
MO000019070	XIAP(h)	XIAP	X-linked inhibitor of apoptosis	356	142.42
MO000020249	26S proteasome(h)	PSMA7, PSMC2, PSMC3, PSMC5, PSMD4, PSMD5	proteasome 20S subunit alpha 7, proteasome 26S subunit, ATPase 2, proteasome 26S subunit, ATPase 3,	374	91.75
MO000018003	PP2A(h)	PPP2CA, PPP2R3A, PPP2R3B, PPP2R5A, PPP2R5B, PPP2R5C, PPP2R5D	protein phosphatase 2 catalytic subunit alpha, protein phosphatase 2 regulatory subunit B''alpha, pr	379	129.52
MO000031189	PKCdelta(h)	PRKCD	protein kinase C delta	390	67.22
MO000019259	c-Cbl(h)	CBL	Cbl proto-oncogene	408	89.08
MO000059577	PKCdelta(h)	PRKCD	protein kinase C delta	411	67.22
MO000019401	Jak1(h)	JAK1	Janus kinase 1	439	137.85

The intracellular regulatory pathways controlled by the above-mentioned master regulators are depicted in Figure 5. This diagram displays the connections between identified transcription factors, which play important roles in the regulation of genes carrying sequence variations, and selected master regulators, which are responsible for the regulation of these TFs.

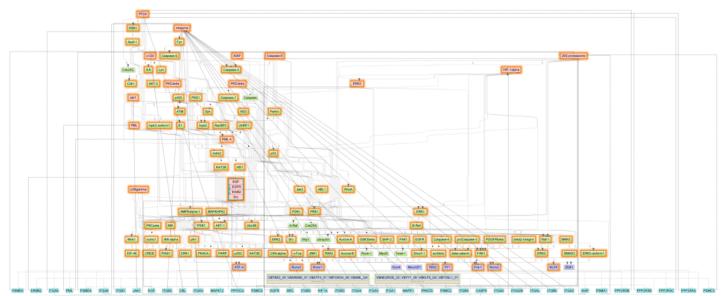


Figure 5. Diagram of intracellular regulatory signal transduction pathways of the most frequently mutated genes in Experiment: short-term survival. Master regulators are indicated by red rectangles, transcription factors are blue rectangles, and green rectangles are intermediate molecules, which have been added to the network during the search for master regulators from selected TFs. Orange frames highlight molecules presented in original mapping.

# 4. Finding prospective drug targets

The identified master regulators that may govern pathology associated genes were checked for druggability potential using HumanPSD™ [5] database of gene-disease-drug assignments and PASS [11-13] software for prediction of biological activities of chemical compounds on the basis of a (Q)SAR approach. Respectively, for each master regulator protein we have computed two Druggability scores: HumanPSD Druggability score and PASS Druggability score. Where Druggability score represents the number of drugs that are potentially suitable for inhibition (or activation) of the corresponding target either according to the information extracted from medical literature (from HumanPSD™ database) or according to cheminformatics predictions of compounds activity against the examined target (from PASS software).

The cheminformatics druggability check is done using a pre-computed database of spectra of biological activities of chemical compounds from a library of all small molecular drugs from HumanPSD™ database, 2507 pharmaceutically active known chemical compounds in total. The spectra of biological activities has been computed using the program PASS [11-13] on the basis of a (Q)SAR approach.

If both Druggability scores were below defined thresholds (see Methods section for the details) such master regulator proteins were not used in further analysis of drug prediction.

As a result we created the following two tables of prospective drug targets (top targets are shown here):



Table 9. Prospective drug targets selected from full list of identified master regulators filtered by Druggability score from HumanPSD™ database. Druggability score contains the number of drugs that are potentially suitable for inhibition (or activation) of the target. The drug targets are sorted according to the Total rank which is the sum of three ranks computed on the basis of the three scores: keynode score, CMA score and expression change score (logFC, if present). See Methods section for details.

### See full table -

Gene symbol	Gene Description	Druggability score	Total rank	Weighted score
ITGAL	integrin subunit alpha L	14	255	191.78
ITGA3	integrin subunit alpha 3	2	255	191.78
ITGB5	integrin subunit beta 5	2	255	191.78
ITGA6	integrin subunit alpha 6	1	255	191.78
ITGA4	integrin subunit alpha 4	5	255	191.78
PSMA7	proteasome 20S subunit alpha 7	2	374	91.75

Table 10. Prospective drug targets selected from full list of identified master regulators filtered by Druggability score predicted by PASS software. Here, the Druggability score for master regulator proteins is computed as a sum of PASS calculated probabilities to be active as a target for various small molecular compounds. The drug targets are sorted according to the Total rank which is the sum of three ranks computed on the basis of the three scores: keynode score, CMA score and expression change score (logFC, if present). See Methods section for details.

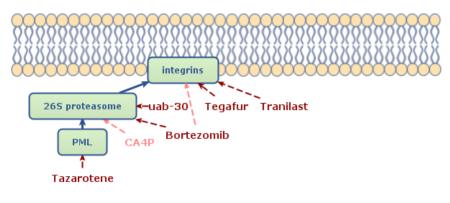
### See full table →

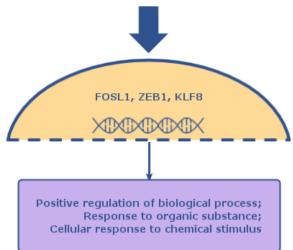
Cana armahal	Come Description	Daving hilliby accus	Takal wasde	Walabaa aaasa
Gene symbol	Gene Description	Druggability score	iotai rank	weignted score
ITGAL	integrin subunit alpha L	0.64	255	191.78
PSMC5	proteasome 26S subunit, ATPase 5	1.28	374	91.75
PSMD5	proteasome 26S subunit, non-ATPase 5	1.28	374	91.75
PSMA7	proteasome 20S subunit alpha 7	2.48	374	91.75
PSMD4	proteasome 26S subunit, non-ATPase 4	1.28	374	91.75
PSMC2	proteasome 26S subunit, ATPase 2	1.28	374	91.75

Below we represent schematically the main mechanism of the studied pathology. In the schema we considered the top two drug targets of each of the two categories computed above. In addition we have added two top identified master regulators for which no drugs may be identified yet, but that are playing the crucial role in the molecular mechanism of the studied pathology. Thus the molecular mechanism of the studied pathology was predicted to be mainly based on the following key master regulators:

- · integrins
- PML
- 26S proteasome

This result allows us to suggest the following schema of affecting the molecular mechanism of the studied pathology:





Drugs which are shown on this schema: Tranilast, Bortezomib, Tazarotene, CA4P, Tegafur and uab-30, should be considered as a prospective research initiative for further drug repurposing and drug development. These drugs were selected as top matching treatments to the most prospective drug targets of the studied pathology, however, these results should be considered with special caution and are to be used for research purposes only, as there is not enough clinical information for adapting these results towards immediate treatment of patients. The drugs given in dark red color on the schema are FDA approved drugs or drugs which have gone through various phases of clinical trials as active treatments against the selected targets.

The drugs given in pink color on the schema are drugs, which were cheminformatically predicted to be active against the selected targets.

# 5. Identification of potential drugs

In the last step of the analysis we strived to identify known activities as well as drugs with cheminformatically predicted activities that are potentially suitable for inhibition (or activation) of the identified molecular targets in the context of specified human diseases(s).

Proposed drugs are top ranked drug candidates, that were found to be active on the identified targets and were selected from 4 categories:

- 1. FDA approved drugs or used in clinical trials drugs for the studied pathology;
- 2. Repurposing drugs used in clinical trials for other pathologies;
- 3. Drugs, predicted by PASS to be active against identified drug targets and against the studied pathology;
- 4. Drugs, predicted by PASS to be active against identified drug targets but for other pathologies.

Proposed drugs were selected on the basis of Drug rank which was computed from the ranks sum based on the individual ranks of the following scores:

- · Target activity score (depends on ranks of all targets that were found for the selected drug);
- Disease activity score (weighted sum of number of clinical trials on disease(s) under study where the selected drug is known to
  be applied or PASS Disease activity score cheminformatically predicted property of the compound to be active against the
  studied disease(s));
- Clinical validity score (applicable only for drugs predicted on the basis of literature curation in HumanPSD™ database (Tables 12 and 13), reflects the number of the highest clinical trials phase on which the drug was tested for any pathology).

You can refer to the Methods section for more details on drug ranking procedure.

Based on the Drug rank, a numerical value of Drug score was calculated, which reflects the potential activity of the respective drug on the overall molecular mechanism of the studied pathology. Drug score values belong to the range from 1 to 100 and are calculated as a quotient of maximum drug rank and the drug rank of the given drug multiplied by 100.

If sufficient information regarding the known associations between predicted drugs and variants identified in the studied pathology was found, this will be reflected in the **Somatic variants** column of the FDA approved and repurposed drugs used in clinical trials tables. Details on these variant-drug associations can be found in the Molecular Tumor Board (MTB) report generated for the studied pathology.

Top drugs of each category are given in the tables below:

# **Drugs approved in clinical trials for Oncology**



Table 11. Clinically approved (FDA, ENA, etc.) drugs for the studied pathology (most promising and clinically approved treatment candidates selected for the identified drug targets on the basis of literature curation in HumanPSD™ database)

See full table →

Name	Target names	Drug score	Disease activity score	Disease trial phase	Approved
Regorafenib	KIT, MAPK1, ABL1, KDR, PDGFRB, FGFR1, BRAF, PDGFRA, RAF1, FRS2, MAPK11, FLT1, AKT1, TEK, MAPK3, RET, AKT2	94	12	Phase 4: Colorectal Neoplasms, Neoplasms, Rectal Neoplasms	Colorectal Neoplasms (FDA)
Fluorouracil	CASP8, CASP3, BAX, ITGAL, FAS, BIRC5, CDKN1A	86	12	Phase 4: Colorectal Neoplasms, Adenocarcinoma, Bowen's Disease, Breast Neoplasms, Carcinoma, Carcinoma, Basal Cell, Carcinoma, Squamous Cell, Cicatrix, Colonic Neoplasms, Digestive System Neoplasms, Esophageal Neoplasms, Foot Diseases, Gastrointestinal Neoplasms, Glaucoma, Glaucoma, Open-Angle, Head and Neck Neoplasms, Hypopigmentation, Intestinal Neoplasms, Keloid, Keratosis, Keratosis, Actinic, Liver Neoplasms, Neoplasm Metastasis, Neoplasms, Neoplasms, Basal Cell, Neoplasms, Second Primary, Neoplasms, Squamous Cell, Pancreatic Neoplasms, Photosensitivity Disorders, Postoperative Complications, Pterygium, Rectal Neoplasms, Recurrence, Skin Diseases, Skin Neoplasms, Squamous Cell Carcinoma of Head and Neck, Stomach Neoplasms, Vitiligo, Warts	Colorectal Neoplasms (ClinicalTrials, ClinicalTrials, ClinicalTrials, ClinicalTrials)
encorafenib	MAPK10, MAPK1, MAPK8, MAPK9, MAPK7, MAPK6, BRAF, MAPK3, MAP2K4, RAF1	86	7	Phase 3: Colorectal Neoplasms, Melanoma, Neoplasms, Rectal Neoplasms	Colorectal Neoplasms (ClinicalTrials, ClinicalTrials, FDA)
Oxaliplatin	CTNNB1, IGF2, CDH1, AKT1, AKT2	76	12	Phase 4: Colorectal Neoplasms, Adenocarcinoma, Colonic Neoplasms, Digestive System Neoplasms, Esophageal Neoplasms, Gastrointestinal Neoplasms, Intestinal Neoplasms, Neoplasm Metastasis, Neoplasms, Neurotoxicity Syndromes, Postoperative Complications, Rectal Neoplasms, Stomach Neoplasms	Colorectal Neoplasms (FDA, PUBMED)
Irinotecan	MAPK10, BIRC5, RUNX3, CDKN1A	63	12	Phase 4: Colorectal Neoplasms, Adenocarcinoma, Carcinoma, Small Cell, Colonic Neoplasms, Esophageal Neoplasms, Gastrointestinal Neoplasms, Lung Neoplasms, Neoplasm Metastasis, Neoplasms, Neuroblastoma, Rectal Neoplasms, Small Cell Lung Carcinoma, Stomach Neoplasms	Colorectal Neoplasms (ClinicalTrials, ClinicalTrials, ClinicalTrials, ClinicalTrials, ClinicalTrials, ClinicalTrials, ClinicalTrials, FDA, FDA)
Bevacizumab	FCGR2A, VEGFA	58	12	Phase 4: Colorectal Neoplasms, Breast Neoplasms, Carcinoma, Carcinoma, Hepatocellular, Carcinoma, Non-Small-Cell Lung, Carcinoma, Ovarian Epithelial, Cataract, Cicatrix, Colonic Neoplasms, Corneal Neovascularization, Diabetic Retinopathy, Dilatation, Pathologic, Edema, Epistaxis, Fallopian Tube Neoplasms, Glaucoma, Hemorrhage, Ischemia, Liver Neoplasms, Lung Neoplasms, Macular Degeneration, Macular Edema, Neoplasm Metastasis, Neoplasms, Neovascularization, Pathologic, Optic Nerve Diseases, Ovarian Neoplasms, Peritoneal Neoplasms, Pterygium, Rectal Neoplasms, Recurrence, Retinal Detachment, Retinal Diseases, Retinal Vein Occlusion, Telangiectasia, Hereditary Hemorrhagic, Telangiectasis, Triple Negative Breast Neoplasms, Vitreous Hemorrhage	Colorectal Neoplasms (FDA, FDA)
Aflibercept	VEGFA	48	7	Phase 3: Colorectal Neoplasms, COVID-19, Choroidal Neovascularization, Diabetes Mellitus, Diabetes Mellitus, Type 1, Diabetes Mellitus, Type 2, Edema, Eye Diseases, Glaucoma, Glaucoma, Neovascular, Macular Degeneration, Macular Edema, Melanoma, Myopia, Myopia, Degenerative, Neoplasm Metastasis, Neoplasms, Neovascularization, Pathologic, Premature Birth, Prostatic Neoplasms, Rectal Neoplasms, Retinal Degeneration, Retinal Diseases, Retinal Vein Occlusion, Retinopathy of Prematurity, Telangiectasis, Vision Disorders, Vision, Low, Wet Macular Degeneration	Colorectal Neoplasms (FDA)
Panitumumab	EGFR	45	12	Phase 4: Colorectal Neoplasms, Colonic Neoplasms, Neoplasms, Rectal Neoplasms	Colorectal Neoplasms (FDA)
Cetuximab	FCGR2A, EGFR	40	12	Phase 4: Colorectal Neoplasms, Carcinoma, Carcinoma, Squamous Cell, Colonic Neoplasms, Head and Neck Neoplasms, Liver Neoplasms, Neoplasm Metastasis, Neoplasms, Rectal Neoplasms, Squamous Cell Carcinoma of Head and Neck	Colorectal Neoplasms (FDA, FDA)
Capecitabine	CDKN2A	25	12	Phase 4: Colorectal Neoplasms, Adenocarcinoma, Brain Neoplasms, Breast Neoplasms, Carcinoma, Colonic Neoplasms, Gastrointestinal Neoplasms,	Colorectal Neoplasms (ClinicalTrials,

The **Disease trial phase** column reflects the maximum clinical trials phase in which the drug was studied for the analyzed pathology.

# **Drugs approved in clinical trials**



Table 12. Drugs used in clinical trials for the studied pathology (most promising treatment candidates selected for the identified drug targets on the basis of literature curation in HumanPSD™ database)

See full table →

Name	Target names	Drug score	Disease activity score	Disease trial phase
Erlotinib	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAPK1, STK11, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BIRC5, BMX, ERBB3, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K1, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, ILK, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, ERBB4, PAK3, BRAF, FER, CAMKK1, TGFBR2, FYN, PRKD2	98	7	Phase 3: Colorectal Neoplasms, Adenocarcinoma, Adenocarcinoma of Lung, Brain Neoplasms, Carcinoma, Carcinoma, Acinar Cell, Carcinoma, Adenosquamous, Carcinoma, Hepatocellular, Carcinoma, Large Cell, Carcinoma, Non-Small- Cell Lung, Carcinoma, Ovarian Epithelial, Carcinoma, Small Cell, Carcinoma, Squamous Cell, Disease Progression, Esophageal Neoplasms, Fallopian Tube Neoplasms, Head and Neck Neoplasms, Lip Neoplasms, Lung Neoplasms, Mouth Neoplasms, Neoplasm Metastasis, Neoplasms, Ovarian Neoplasms, Pancreatic Intraductal Neoplasms, Pancreatic Neoplasms, Peritoneal Neoplasms, Rectal Neoplasms, Small Cell Lung Carcinoma, Squamous Cell Carcinoma of Head and Neck, Thoracic Neoplasms
Sunitinib	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, CAMKK1, SIRT1, TGFBR2, FYN, PRKD2	97	7	Phase 3: Colorectal Neoplasms, Adenoma, Adenoma, Islet Cell, Brain Neoplasms, Breast Neoplasms, Carcinoma, Carcinoma, Hepatocellular, Carcinoma, Islet Cell, Carcinoma, Non-Small-Cell Lung, Carcinoma, Renal Cell, Gastrointestinal Stromal Tumors, Glioblastoma, Kidney Diseases, Kidney Neoplasms, Liver Neoplasms, Lung Neoplasms, Neoplasms, Neoplasms by Site, Pancreatic Neoplasms, Prostatic Neoplasms, Rectal Neoplasms, Recurrence, Urogenital Neoplasms, Urologic Neoplasms
Gefitinib	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, IGF2, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAKA3, BRAF, FER, CAMKK1, TGFBR2, FYN, PRKD2	97	7	Phase 3: Colorectal Neoplasms, Adenocarcinoma, Adenocarcinoma of Lung, Adenocarcinoma, Bronchiolo-Alveolar, Brain Neoplasms, Breast Neoplasms, Carcinoma, Carcinoma, Non-Small-Cell Lung, Carcinoma, Squamous Cell, Head and Neck Neoplasms, Lung Neoplasms, Neoplasm Metastasis, Neoplasms, Rectal Neoplasms, Squamous Cell Carcinoma of Head and Neck, Thoracic Neoplasms, Urinary Bladder Neoplasms

Lapatinib

ITGA5, VEGFA, ITGB1, MTOR, ITGB4, FGF2, 96 ITGA3, ITGA6, ITGB5, EGFR, PTK2, CASP3, AKT1, ITGB3, AKT2 Phase 3: Colorectal Neoplasms, Adenocarcinoma, Ascites, Breast Neoplasms, Carcinoma, Cholangiocarcinoma, Colonic Neoplasms, Esophageal Neoplasms, Head and Neck Neoplasms, Liver Neoplasms, Nasopharyngeal Carcinoma, Neoplasm Metastasis, Neoplasms, Pancreatic Neoplasms, Rectal Neoplasms, Paccurrence, Stomach Neoplasms, Lirinary, Bladder Neoplasms

BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7 MAPK11, FLT1, PRKCD, LYN, CSNK2A1 AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BIRC5, BMX, PIM2, RIPK2, FGFR3 RPS6KA2, PAK4, NTRK2, PIM3, YES1 NUAK1, CAMK2D, ITK, MAP3K4, PAK1 CLK1, CDK5, RET, ABL2, CSF1R, STK10, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, MAP2K2, MAP2K3, RPS6KA5, CSK PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES PLK4, DAPK3, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, CAMKK1, TGFBR2, FYN, PRKD2 Recurrence, Stomach Neoplasms, Urinary Bladder Neoplasms Phase 2: Colorectal Neoplasms, Adenocarcinoma, Adenoma, Astrocytoma, Bile Duct Neoplasms, Brain Neoplasms, Breast Diseases, Breast Neoplasms, Breast Neoplasms, Male, Carcinoma, Carcinoma, Acinar Cell, Carcinoma, Adenoid Cystic, Carcinoma, Ductal, Carcinoma, Ductal, Breast, Carcinoma, Hepatocellular, Carcinoma, Mucoepidermoid, Carcinoma, Non-Small-Cell Lung, Carcinoma, Ovarian Epithelial, Carcinoma, Small Cell, Carcinoma, Squamous Cell, Carcinoma, Transitional Cell, Carcinoma, Verrucous, Central Nervous System Neoplasms, Cholangiocarcinoma, Cysts, Dermoid Cyst, Diarrhea, Digestive System Neoplasms, Endometrial Neoplasms, Ependymoma, Esophageal Neoplasms, Esophageal Squamous Cell Carcinoma, Fibroma, Gallbladder Neoplasms, Gastrointestinal Neoplasms, Glioblastoma, Glioma, Gliosarcoma, Head and Neck Neoplasms, Hypopharyngeal Neoplasms, Inflammatory Breast Neoplasms, Intestinal Neoplasms, Laryngeal Diseases, Laryngeal Neoplasms, Liver Neoplasms, Lung Neoplasms, Medulloblastoma, Melanoma, Mucoepidermoid Tumor, Nasopharyngeal Carcinoma, Neoplasm Metastasis, Neoplasms, Neoplasms, Second Primary, Neoplasms, Unknown Primary, Neoplastic Cells, Circulating, Nervous System Neoplasms, Neurilemmoma, Neuroblastoma, Neurofibroma, Neurofibromatoses, Neurofibromatosis 1, Neurofibromatosis 2, Neuroma, Neuroma, Acoustic, Oligodendroglioma, Oropharyngeal Neoplasms, Ovarian Neoplasms, Pancreatic Neoplasms, Paranasal Sinus Neoplasms, Peritoneal Neoplasms, Pharyngeal Neoplasms, Pituitary Diseases, Pituitary Neoplasms, Prolactinoma, Prostatic Neoplasms, Prostatic Neoplasms, Castration-Resistant, Rectal Neoplasms, Recurrence, Salivary Gland Neoplasms, Sarcoma, Small Cell Lung Carcinoma, Spinal Cord Neoplasms, Squamous Cell Carcinoma of Head and Neck, Stomach Neoplasms, Thymoma, Tongue Neoplasms, Urinary Bladder Neoplasms, Uterine Cervical Neoplasms, Uterine Neoplasms

The **Disease trial phase** column reflects the maximum clinical trials phase in which the drug was studied for the analyzed pathology.

7

5

95

# Repurposing drugs



Table 13. Repurposed drugs used in clinical trials for other pathologies (prospective drugs against the identified drug targets on the basis of literature curation in  $HumanPSD^{TM}$  database)

Name	Target names	Drug score	Maximum trial phase
seliciclib	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, CDK4, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, PRKCZ, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, CHEK2, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, HIPK2, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, ROCK2, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, SGK1, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, IKBKB, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, CDK1, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMY11, RPS6KB1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, MAPK13, CAMKK1, TGFBR2, FYN, PRKD2	81	Phase 2: ACTH-Secreting Pituitary Adenoma, Adenoma, Carcinoma, Non-Small-Cell Lung, Cystic Fibrosis, Cysts, Fibrosis, Pituitary ACTH Hypersecretion, Pituitary Neoplasms
1-(5-Tert- Butyl-2-P- Tolyl-2h- Pyrazol-3- YI)-3-[4-(2- Morpholin-4- YI-Ethoxy)- Naphthalen- 1-YI]-Urea	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, PRKCZ, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, CHEK2, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, HIPK2, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, ROCK2, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, SGK1, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, IKBKB, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, RPS6KB1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, MAPK13, CAMKK1, TGFBR2, FYN, PRKD2	80	Phase 2: Arthritis, Arthritis, Rheumatoid, Psoriasis
ruboxistaurin	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, PRKCZ, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, CHEK2, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, HIPK2, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, ROCK2, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, SGK1, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, IKBKB, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, RPS6KB1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, MAPK13, CAMKK1, TGFBR2, FYN, PRKD2	80	Phase 3: Diabetes Mellitus, Diabetes Mellitus, Type 1, Diabetes Mellitus, Type 2, Diabetic Neuropathies, Diabetic Retinopathy, Edema, Macular Edema, Nervous System Diseases, Peripheral Nervous System Diseases, Retinal Diseases
Tofacitinib	BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN, CSNK2A1, AKT2, STK3, PDPK1, MAPK10, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1,	80	Phase 4: Alopecia, Alopecia Areata, Aortic Arch Syndromes, Arteritis, Arthritis, Arthritis, Psoriatic, Arthritis, Rheumatoid, COVID-19, Colitis, Colitis, Ulcerative, Disease, Embolism, Granuloma, Granulomatosis with Polyangiitis, Infections, Lung Diseases, Lung Diseases, Interstitial, Necrosis, Rheumatic Fever, ST Elevation Myocardial Infarction, Spondylarthritis, Spondylitis, Spondylitis, Ankylosing, Systemic Vasculitis, Takayasu Arteritis, Thromboembolism, Ulcer, Vasculitis

CAMK2D, ITK, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, ROCK2, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, SGK1, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, RPS6KA3, CAMK2G, MET, PRKCQ, EPHA4, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, RPS6KB1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, MAPK13, CAMKK1, TGFBR2, FYN, PRKD2

BMPR1A, IKBKE, ABL1, NEK7, JAK3, PAK2, PRKACA, GSK3B, MAP3K11, CDK4, SYK, NEK6, EIF2AK2, ERBB2, MAPK3, MAP2K6, JAK1, MELK, MAPK8, SRC, CAMKK2, PRKD3, CSNK1G2, MAP4K1, STK11, CSNK1G1, CLK4, MAPK12, PLK3, WEE1, MAPK7, MAPK11, FLT1, PRKCD, LYN CSNK2A1, AKT2, STK3, PDPK1, MAPK10, CDK6, KDR, MAPK6, PRKCA, CSNK1E, PDGFRA, RAF1, AURKB, CDK7, MAPK14, CSNK1D, CHEK1, PTK6, BMX, PIM2, RIPK2, FGFR3, RPS6KA2, PAK4, NTRK2, PIM3, YES1, NUAK1, CAMK2D, ITK, XIAP, MAP3K4, PAK1, CLK1, CDK5, RET, ABL2, CSF1R, STK10, MAP4K4, MARK3, MAP3K10, SLK, INSR, NEK2, MAP2K2, MAP2K3, RPS6KA5, CSK, PRKD1, TEK, PKN1, TYK2, IGF1R, MAP3K5, CSNK2A2, PDGFRB, MAP2K1, MAP4K3, STK4, CAMK2A, PTK2, MAPKAPK2, FES, PLK4, DAPK3, RPS6KA3 CAMK2G, MET, PRKCQ, EPHA4, CDK1, MAP2K4, PRKAA1, RPS6KA1, EGFR, PLK1, PRKCH, PTK2B, AKT1, AURKA, JAK2, MAP3K20, PKMYT1, KIT, MAPK1, DYRK1B, EPHB2, MAPK9, GSK3A, MERTK, PRKCE, FGFR1, PAK3, BRAF, FER, CAMKK1, TGFBR2, FYN, PRKD2

Phase 2: Adenocarcinoma, Brain Abscess, Breast Neoplasms, Carcinoma, Hepatocellular, Carcinoma, Ovarian Epithelial, Carcinoma, Renal Cell, Embolism, Endometrial Neoplasms, Esophageal Neoplasms, Germinoma, Granuloma, Head and Neck Neoplasms, Hodgkin Disease, Hypereosinophilic Syndrome, Immunoblastic Lymphadenopathy, Kidney Neoplasms, Leukemia, Leukemia, Basophilic, Acute, Leukemia, Eosinophilic, Acute, Leukemia, Erythroblastic, Acute, Leukemia, Lymphocytic, Chronic, B-Cell, Leukemia, Lymphoid, Leukemia, Megakaryoblastic, Acute, Leukemia, Monocytic, Acute, Leukemia, Myeloid, Leukemia, Myeloid, Acute, Leukemia, Myelomonocytic, Acute, Leukemia, Prolymphocytic, Leukemia, T-Cell, Leukemia-Lymphoma, Adult T-Cell, Liver Neoplasms, Lymphadenopathy, Lymphatic Diseases, Lymphoma, Lymphoma, B-Cell, Lymphoma, B-Cell, Marginal Zone, Lymphoma, Follicular, Lymphoma, Large B-Cell, Diffuse, Lymphoma, Large-Cell, Anaplastic, Lymphoma, Mantle-Cell, Lymphoma, Non-Hodgkin, Lymphoma, T-Cell, Lymphoma, T-Cell, Cutaneous, Lymphomatoid Granulomatosis, Melanoma, Multiple Myeloma, Mycoses, Mycosis Fungoides, Myelodysplastic Syndromes, Neoplasms, Neoplasms, Germ Cell and Embryonal, Neoplasms, Plasma Cell, Ovarian Neoplasms, Pancreatic Neoplasms, Peritoneal Neoplasms, Precursor Cell Lymphoblastic Leukemia-Lymphoma, Preleukemia, Prostatic Neoplasms, Prostatic Neoplasms, Castration-Resistant, Recurrence, Sarcoma, Seminoma, Sezary Syndrome, Stomach Neoplasms, Testicular Neoplasms, Thromboembolism, Waldenstrom Macroglobulinemia

The Maximum trial phase column reflects the maximum clinical trials phase in which the drug was studied for any pathology.

80



Flavopiridol

Table 14. Prospective drugs, predicted by PASS software to be active against the identified drug targets with predicted activity against the studied disease(s) (drug candidates predicted with the cheminformatics tool PASS)

	See	full	table	-
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Name	Target names	Drug score	Target activity score
Camptothecin	HIF1A, CASP3	80	0.31
Topotecan	HIF1A, CASP3	80	0.31
LE-SN38	HIF1A, CASP3	80	0.29
Ouabain	STAT3, CASP8, HIF1A, CASP3, STAT1, RELA	76	0.19
Eucalyptol	VEGFA, TLR3, TLR4, KDR, FLT1	74	0.17



Table 15. Prospective drugs, predicted by PASS software to be active against the identified drug targets, though without cheminformatically predicted activity against the studied disease(s) (drug candidates predicted with the cheminformatics tool PASS)

See full table  $\rightarrow$ 

Name	Target names	Drug score	Target activity score
2,6-Dihydroanthra/1,9-Cd/Pyrazol-6-One	MAPK10, RPS6KA3, IRAK4, CDK6, CAMK2G, CSNK1A1, STAT1, PAK2, EPHA4, CSNK1E, CDK1, MAP2K4, CDK4, AURKB, CDK7, PRKAA1, HSP90AA1, CSNK1D, SGK1, MAP2K3, PTK2B, AKT1, AURKA, CHEK1, PKN1, MAP2K6, MAPK8, MAPK9, EPHB2, DYRK1B, GSK3A, PAK4, PRKD3, CSNK1G2, PAK3, CLK4, CSNK1G1, CAMK2D, CAMK2A, PAK1, CLK1, IRAK1, CDK5, CHEK2, FES, AKT2, DAPK3	96	12.11
Iodophenyl	STK10, ROCK2, MAP4K4, MARK3, NEK7, PAK2, GSK3B, SLK, VRK1, NEK2, SGK1, NEK6, EIF2AK2, TAOK2, RPS6KA5, MELK, CSNK2A2, CSNK1G2, MAP4K1, TRIO, STK11, MAP4K3, CSNK1G1, STK4, NEK10, MAP4K2, MAPKAPK2, IRAK1, CSNK2A1, CHEK2, AKT2, PDPK1, DAPK3, STK3, RPS6KA3, PIK3CG, IRAK4, PINK1, CSNK1A1, CSNK1E, RAF1, PRKAA1, RPS6KA1, AURKB, ILK, CSNK1D, MASTL, HIPK1, AKT1, AURKA, CHEK1, ATM, LMTK2, PKMYT1, RPS6KB1, TBK1, ATR, RIPK2, PIM2, HIPK2, MTOR, GSK3A, RPS6KA2, PAK4, PIM3, PRKDC, NUAK1, PAK3, PASK, BRAF, PAK1, CDK5	96	19.8
Rbt205 Inhibitor	RPS6KA3, CDK6, CAMK2G, GRK2, MAP3K10, PRKCQ, PRKACA, GSK3B, PRKCA, CDK1, MAP2K4, CDK4, CDK7, RPS6KA1, PRKAA1, SGK1, PRKCH, PRKD1, PKN1, MAP2K6, RPS6KA2, GSK3A, GRK5, PRKD3, PRKCE, PRKCZ, CAMK2D, PTK2, CAMK2A, SIRT1, PRKCD, PRKCI, CDK5, DAPK3	95	19.7
Uracil mustard	MET, KDR, ABL1, JAK3, EPHA4, PDGFRA, INSR, MST1R, EGFR, SYK, PTK2B, EIF2AK2, ERBB2, PTK6, JAK2, TEK, CSK, TYK2, MELK, JAK1, IGF1R, KIT, BMX, RIPK2, ERBB3, EPHB2, FGFR3, SRC, MTOR, NTRK2, MERTK, PDGFRB, YES1, FGFR1, ERBB4, FER, ITK, WEE1, PTK2, FLT1, PRKCD, LYN, FYN, RET, FES, CSF1R, ABL2	94	6.65
7-[4- (Dimethylamino)Phenyl]- N-Hydroxy-4,6- Dimethyl-7-Oxo-2,4- Heptadienamide	HDAC4, HDAC2, HDAC3, HDAC1	93	3.2

As the result of drug search we propose the following drugs as most promising candidates for treating the pathology under study: Erlotinib, seliciclib, Camptothecin and 2,6-Dihydroanthra/1,9-Cd/Pyrazol-6-One. These drugs were selected for acting on the following targets: DYRK1B and HIF1A, which were predicted to be active in the molecular mechanism of the studied pathology.

The selected drugs are top ranked drug candidates from each of the four categories of drugs: (1) FDA approved drugs or used in clinical trials drugs for the studied pathology; (2) repurposing drugs used in clinical trials for other pathologies; (3) drugs, predicted by PASS software to be active against the studied pathology; (4) drugs, predicted by PASS software to be repurposed from other pathologies.

# Supplementary drug info

In addition to the approved and repurposed drugs proposed by Genome Enhancer, below the **Supplementary drug info** table is given, which contains an extended list of drugs used for treatment of neoplasms. Those drugs which were predicted by Genome Enhancer as prospective treatment candidates for the studied case (both approved and repurposed) have a respective **Predicted Drug Score** assigned to them. This value on a scale from 1 to 100 reflects the potential activity of the respective drug on the overall molecular mechanism of the studied pathology. The **Predicted Drug Score** column contains the "-" (Not Identified) value in case the drug targets of the respective treatment were not found in the molecular mechanism of the studied pathology.

Table 16. Supplementary drug info: extended list of drugs used for treatment of neoplasms with respective drug scores predicted for the studied pathology.

Drug	Disease	Predicted Drug Score	Somatic variants
Abarelix	Prostatic Neoplasms	-	
Abemaciclib	Breast Neoplasms	52	
Abiraterone	Prostatic Neoplasms, Castration-Resistant	-	
Abiraterone acetate	Prostatic Neoplasms, Castration-Resistant	-	
Acalabrutinib	Lymphoma, Mantle-Cell	30	
Acitretin	Psoriasis	27	
Ado-trastuzumab emtansine	Breast Neoplasms   Neoplasms	78	
Afatinib	Carcinoma, Non-Small-Cell Lung	73	
Aflibercept	Colorectal Neoplasms Diabetic Retinopathy Edema Vascular Diseases Wet Macular Degeneration	48	
Alectinib	Carcinoma, Non-Small-Cell Lung	40	
Alemtuzumab	Brain Abscess Leukemia, Lymphocytic, Chronic, B-Cell Multiple Sclerosis Multiple Sclerosis, Relapsing-Remitting Sclerosis	-	
Alitretinoin	Sarcoma, Kaposi	-	
Alpelisib	Breast Neoplasms	-	
Altretamine	Ovarian Neoplasms	-	
Aminolevulinic acid	Keratosis Keratosis, Actinic	-	
Anagrelide	Thrombocythemia, Essential Thrombocytosis	-	
Anastrozole	Breast Neoplasms Hypersensitivity Obesity Obesity, Morbid Recurrence Weight Loss	21	
Apalutamide	Prostatic Neoplasms, Castration-Resistant	-	
Aprepitant	Nausea Neoplasms Postoperative Nausea and Vomiting	-	
Arsenic trioxide	Leukemia, Promyelocytic, Acute	66	
Atezolizumab	Carcinoma, Non-Small-Cell Lung Carcinoma, Transitional Cell Triple Negative Breast Neoplasms	-	
Avelumab	Carcinoma, Merkel Cell Carcinoma, Renal Cell Carcinoma, Transitional Cell	-	
Axitinib	Carcinoma, Renal Cell	87	
Azacitidine	Anemia, Refractory Anemia, Refractory, with Excess of Blasts Leukemia, Myelomonocytic, Chronic Myelodysplastic Syndromes Preleukemia Syndrome	48	
Belinostat	Lymphoma, T-Cell, Peripheral	65	
Bendamustine	Leukemia, Lymphocytic, Chronic, B-Cell Leukemia, Lymphoid	-	
Bevacizumab	Breast Neoplasms Colonic Neoplasms Colorectal Neoplasms Corneal Neovascularization Diabetic Retinopathy Dilatation, Pathologic Edema Epistaxis Glaucoma Hemorrhage Macular Degeneration Macular Edema Neoplasm Metastasis Neoplasms Neovascularization, Pathologic Optic Nerve Diseases Pterygium Rectal Neoplasms Retinal Detachment Retinal Diseases Retinal Vein Occlusion Telangiectasia, Hereditary Hemorrhagic Telangiectasis Vitreous Hemorrhage	58	
Bexarotene	Lymphoma, T-Cell Lymphoma, T-Cell, Cutaneous	3	
Bicalutamide	Prostatic Neoplasms	7	
Binimetinib	Melanoma	77	
Blinatumomab	Precursor B-Cell Lymphoblastic Leukemia-Lymphoma	-	
Bortezomib	Brain Abscess Glomerulonephritis Glomerulonephritis, IGA Kidney Diseases Multiple Myeloma Neoplasms, Plasma Cell Nephritis Renal Insufficiency	66	
Bosutinib	Leukemia, Myelogenous, Chronic, BCR-ABL Positive	74	
Brentuximab vedotin	Hodgkin Disease Lymphoma Lymphoma, Large-Cell, Anaplastic Lymphoma, T-Cell, Peripheral	-	
Brigatinib	Carcinoma, Non-Small-Cell Lung	49	
Buserelin	Prostatic Neoplasms	-	
Cabazitaxel	Prostatic Neoplasms, Castration-Resistant	78	
Cabergoline	Drug-Related Side Effects and Adverse Reactions Pituitary	3	
Cabergonne	Neoplasms		

Capecitabine	Breast Neoplasms Colonic Neoplasms Colorectal Neoplasms	25	
	Carcinoma, Non-Small-Cell Lung Lung		
Carboplatin	Neoplasms Neoplasms Neuroendocrine Tumors Ovarian	-	XRCC1:Q399R:sensitivity/response:B2
	Neoplasms Retinoblastoma		
Carfilzomib	Multiple Myeloma	63	
Carmustine	Astrocytoma Glioblastoma Hodgkin	-	
Caultinib	Disease Medulloblastoma Multiple Myeloma Neoplasms	75	
Ceritinib	Carcinoma, Non-Small-Cell Lung	75 40	
Cetuximab	Colorectal Neoplasms	40	
	Anemia Calcinosis Cardiovascular Diseases Hyperparathyroidism Hyperparathyroidism,		
Cinacalcet	Secondary Kidney Diseases Kidney Failure, Chronic Neoplasm		
Ciliacaicet	Metastasis Neoplasms Parathyroid Neoplasms Renal	-	
	Insufficiency Vascular Calcification Vascular Diseases Vision		
	Disorders  Considers  Considers  Considers  Considers		
Cisplatin	Carcinoma, Squamous Cell Neoplasms Uterine Cervical Neoplasms Carcinoma, Non-Small-Cell Lung Esophageal	80	XRCC1:Q399R:sensitivity/response:B2
Cispidin	Neoplasms Carcinoma	00	ARCC1.Q333R.3eli3itivity/response.b2
Cladribine	Leukemia, Hairy Cell	48	
Clofarabine	Precursor Cell Lymphoblastic Leukemia-Lymphoma	38	
Cobimetinib	Melanoma	82	
Copanlisib	Lymphoma, Follicular	79	
Crizotinib	Carcinoma, Non-Small-Cell Lung	90	
Cyproterone acetate	Prostatic Neoplasms	-	
Dabrafenib	Melanoma	40	
Dacomitinib	Carcinoma, Non-Small-Cell Lung	84	
Daratumumab	Multiple Myeloma	32	
	Leukemia, Myelogenous, Chronic, BCR-ABL Positive Leukemia,		
Dasatinib	Myeloid, Chronic-Phase Precursor Cell Lymphoblastic Leukemia-	92	
	Lymphoma		
S	Anemia, Refractory   Anemia, Refractory, with Excess of		
Decitabine	Blasts Leukemia, Myelomonocytic, Chronic Myelodysplastic	13	
Describe	Syndromes  Condition Process   Proceedings   Nacral Consulty   Process   Pro	Г1	
Degarelix	Cardiovascular Diseases   Prostatic Neoplasms   Vascular Diseases	51	
	Arthritis, Rheumatoid Bone Diseases Bone Diseases, Metabolic Breast		
5	Neoplasms Hyperparathyroidism Hyperparathyroidism,		
Denosumab	Primary Metabolic Diseases Neoplasm	-	
	Metastasis Neoplasms Osteoporosis Osteoporosis,		
_	Postmenopausal Prostatic Neoplasms		
Dexrazoxane	Breast Neoplasms   Cardiomyopathies	5	
Dienogest	Menorrhagia	-	
Dinutuximab	Neuroblastoma	-	
Docetaxel	Breast Neoplasms Carcinoma, Non-Small-Cell Lung Prostatic Neoplasms Squamous Cell Carcinoma of Head and Neck Stomach	44	
Docetaxei	Neoplasms	44	
	Neoplasms Multiple Myeloma Carcinoma, Ovarian		
Doxorubicin	Epithelial Ovarian Neoplasms Leukemia, Lymphoid Breast	80	
DOXOFUDICIT	Neoplasms Lymphoma, Follicular Thyroid Neoplasms Triple	80	
	Negative Breast Neoplasms Glioma		
Durvalumab	Carcinoma, Non-Small-Cell Lung Carcinoma, Transitional Cell	-	
Dutasteride	Alcoholism Hyperplasia Hypertrophy Neoplasms Prostatic	-	
D 11.11	Hyperplasia		
Duvelisib	Leukemia, Lymphocytic, Chronic, B-Cell Lymphoma, Follicular	59	
Elotuzumab	Multiple Myeloma	70	
Enasidenib	Leukemia, Myeloid, Acute	-	
Encorafenib	Colorectal Neoplasms Melanoma	86	
Enfortumab vedotin	Carcinoma, Transitional Cell Neoplasms		
Entrectinib	Carcinoma, Non-Small-Cell Lung	50	
Enzalutamide	Prostatic Neoplasms   Prostatic Neoplasms, Castration-Resistant	-	
Epirubicin	Breast Neoplasms	57	
Erdafitinib	Urinary Bladder Neoplasms	81	
Eribulin	Breast Neoplasms Drug-Related Side Effects and Adverse Reactions Neoplasms	-	
	Carcinoma, Non-Small-Cell Lung Neoplasms Pancreatic		
Erlotinib	Neoplasms	98	
Erlotinib	<u> </u>		
hydrochloride	Carcinoma, Non-Small-Cell Lung Gastrointestinal Stromal Tumors	-	
Estramustine	Prostatic Neoplasms	11	
Ethinyl Estradiol	Acne Vulgaris Neoplasms	4	
	Angiomyolipoma Arthrogryposis Astrocytoma Breast		
	Neoplasms   Carcinoma, Renal Cell   Cysts   Idiopathic Pulmonary	0.2	
Everolimus	Fibrosis Kidney Diseases, Cystic Kidney Failure,	83	
	Chronic Lipoma Neuroendocrine Tumors Primary Graft Dysfunction Sclerosis Tuberous Sclerosis		
Exemestane	Breast Neoplasms	-	
Fedratinib	Primary Myelofibrosis	55	
Finasteride	Hyperplasia Neoplasms Prostatic Hyperplasia	33	
Flavopiridol	Leukemia, Lymphocytic, Chronic, B-Cell	80	
	Skin Neoplasms Neoplasms, Basal Cell Neoplasms, Second		
Fluorouracil	Primary Neoplasms, Squamous Cell Neoplasms Colorectal	86	
	Neoplasms Pancreatic Neoplasms		
	Neopiasitis paricieatic Neopiasitis		

Fluoxymesterone	Breast Neoplasms Hypogonadism Puberty, Delayed	12
Flutamide	Premenstrual Dysphoric Disorder Premenstrual Syndrome Prostatic Neoplasms	42
Fulvestrant	Breast Neoplasms	61
Gefitinib	Carcinoma, Non-Small-Cell Lung	97
	Breast Neoplasms Carcinoma, Non-Small-Cell Lung Ovarian	
Gemcitabine	Neoplasms Pancreatic Neoplasms	83
Gemtuzumab ozogamicin	Leukemia, Myeloid, Acute	6
Gilteritinib	Leukemia, Myeloid, Acute	63
Glasdegib	Leukemia, Myeloid, Acute	-
Goserelin	Atrophy Breast Neoplasms Bulbo-Spinal Atrophy, X- Linked Endometriosis Muscular Atrophy Myoma Prostatic Neoplasms	-
Histrelin	Puberty, Precocious	-
Homoharringtonine	Leukemia, Myelogenous, Chronic, BCR-ABL Positive	72
Ibritumomab	Lymphoma, B-Cell Lymphoma, Follicular	6
Ibrutinib	Graft vs Host Disease Leukemia, Lymphocytic, Chronic, B-Cell Lymphoma, B-Cell, Marginal Zone Lymphoma, Mantle-Cell Waldenstrom Macroglobulinemia	87
Idarubicin	Leukemia, Myeloid, Acute	39
Idelalisib	Leukemia, Lymphocytic, Chronic, B-Cell Lymphoma, Follicular	63
Ifosfamide	Neoplasms	-
Imatinib	Leukemia, Myelogenous, Chronic, BCR-ABL Positive Mastocytosis, Systemic Neoplasms	91
Inotuzumab ozogamicin	Precursor B-Cell Lymphoblastic Leukemia-Lymphoma	-
Ipilimumab	Carcinoma, Renal Cell Melanoma	38
Irinotecan	Colorectal Neoplasms	63
Ivosidenib	Leukemia, Myeloid, Acute	-
Ixabepilone	Breast Neoplasms	-
Ixazomib	Multiple Myeloma	-
Lapatinib	Breast Neoplasms	95
Larotrectinib	Neoplasm Metastasis	56
Lenalidomide	Brain Abscess Lupus Erythematosus, Cutaneous Myelodysplastic Syndromes Neoplasms, Plasma Cell	23
Lenvatinib	Carcinoma, Hepatocellular Carcinoma, Renal Cell Thyroid	84
Letrozole	Neoplasms Breast Neoplasms Cysts Fibroma Myofibroma Myoma Ovarian	22
Leuprolide	Cysts Syndrome Hot Flashes Ovarian Hyperstimulation Syndrome Prostatic	-
	Neoplasms Puberty, Precocious	
Levamisole	Ascariasis Colonic Neoplasms Helminthiasis	-
Levonorgestrel Lomustine	Epilepsy Hyperplasia Menorrhagia Brain Neoplasms Hodgkin Disease	-
Lonafarnib	Breast Neoplasms Carcinoma, Non-Small-Cell Lung Central Nervous System Neoplasms Colorectal Neoplasms Gliosarcoma Head and Neck Neoplasms Leukemia, Myelomonocytic, Chronic  Liver Neoplasms Lymphoma Myelodysplastic Syndromes Ovarian Neoplasms Urethral Neoplasms Urinary Bladder Neoplasms	38
Lorlatinib	Carcinoma, Non-Small-Cell Lung	65
Masoprocol	Keratosis, Actinic	16
Medroxyprogesterone Acetate	Depression Depression, Postpartum Depressive Disorder Metrorrhagia Neoplasms Uterine Hemorrhage	21
Megestrol acetate	Acquired Immunodeficiency Syndrome Bites and Stings Breast Neoplasms Pain Wasting Syndrome	-
Methotrexate	Neoplasms Breast Neoplasms Head and Neck Neoplasms Ovarian Neoplasms Lymphoma, T-Cell, Peripheral Brain Neoplasms Colorectal Neoplasms Neuroblastoma Carcinoma, Squamous Cell	49
Methyltestosterone	Breast Neoplasms Hypogonadism Puberty, Delayed	-
Midostaurin	Leukemia, Mast-Cell Leukemia, Myeloid, Acute Mastocytosis,	79
	Systemic	
Mitotane	Adrenocortical Carcinoma	-
Mitoxantrone	Autoimmune Diseases Autoimmune Diseases of the Nervous System Demyelinating Autoimmune Diseases, CNS Immune System Diseases Leukemia, Myeloid, Acute Multiple Sclerosis Myelitis Myelitis, Transverse Nervous System Diseases Neuromyelitis Optica Prostatic Neoplasms, Castration-Resistant	34
Mogamulizumab	Mycosis Fungoides Neoplasms Sezary Syndrome	-
Moxetumomab	Leukemia, Hairy Cell Neoplasms	21
pasudotox		<u>-1</u>
Necitumumab	Carcinoma, Non-Small-Cell Lung Neoplasms	-
Nelarabine	Precursor T-Cell Lymphoblastic Leukemia-Lymphoma	-
Neratinib	Breast Neoplasms	83
Nilotinib	Blast Crisis Leukemia, Myelogenous, Chronic, BCR-ABL Positive Leukemia, Myeloid, Chronic-Phase	72
waster and the second second		
Nilutamide Nintedanib	Prostatic Neoplasms Fibrosis Idiopathic Pulmonary Fibrosis	- 91

Niraparib	Carcinoma, Ovarian Epithelial Fallopian Tube Neoplasms Peritoneal Neoplasms	45
Nivolumab	Carcinoma, Non-Small-Cell Lung Kidney Neoplasms Neoplasms Lung Neoplasms Melanoma	-
Obinutuzumab	Leukemia, Lymphocytic, Chronic, B-Cell	<u>-</u>
Octreotide	Acromegaly Adenoma Ascites Carcinoid Tumor Fistula Pancreatic	52
	Fistula Pituitary Diseases Renal Insufficiency Vipoma	52
Ofatumumab	Leukemia, Lymphocytic, Chronic, B-Cell	-
Olaparib	Breast Neoplasms Carcinoma, Ovarian Epithelial Fallopian Tube Neoplasms Ovarian Neoplasms Pancreatic Neoplasms Peritoneal Neoplasms Prostatic Neoplasms, Castration-Resistant	43
Olaratumab	Sarcoma	-
Osimertinib	Carcinoma, Non-Small-Cell Lung	85
Oxaliplatin	Colonic Neoplasms Colorectal Neoplasms Neoplasms Rectal Neoplasms	76
Paclitaxel	Acute Coronary Syndrome Angina Pectoris Arteriosclerosis Breast Neoplasms Carcinoma, Non-Small-Cell Lung Cardiovascular Diseases Coronary Artery Disease Coronary Disease Coronary Stenosis Heart Diseases Myocardial Ischemia Ovarian Neoplasms Vascular Diseases	79
Palbociclib	Breast Neoplasms	46
Panitumumab	Colorectal Neoplasms	45
Panobinostat	Multiple Myeloma	63
Pazopanib	Carcinoma Carcinoma, Renal Cell Sarcoma	81
Pembrolizumab	Carcinoma, Hepatocellular Carcinoma, Merkel Cell Carcinoma, Non-Small-Cell Lung Carcinoma, Renal Cell Carcinoma, Transitional Cell Hodgkin Disease Melanoma Neoplasms Stomach Neoplasms	-
Pemetrexed	Carcinoma, Non-Small-Cell Lung Mesothelioma	-
Pentostatin	Leukemia, Hairy Cell	11
Pertuzumab	Breast Neoplasms	81
Pomalidomide	Multiple Myeloma	11
Ponatinib	Leukemia, Myelogenous, Chronic, BCR-ABL Positive Precursor Cell Lymphoblastic Leukemia-Lymphoma	76
Pralatrexate	Lymphoma, T-Cell, Peripheral	-
Radium Ra 223 Dichloride	Prostatic Neoplasms, Castration-Resistant	-
Ramucirumab	Stomach Neoplasms	-
Rasburicase	Hyperuricemia Leukemia Lymphoma Neoplasms Syndrome Tumor Lysis Syndrome	-
Regorafenib	Colorectal Neoplasms	94
Relugolix	Prostatic Neoplasms	-
Ribociclib	Breast Neoplasms	52
Rituximab	Arthritis Arthritis, Rheumatoid Granulomatosis with Polyangiitis Leukemia Leukemia, Lymphoid Lymphoma, Lymphoid Lymphoma, Follicular Lymphoma, Non-Hodgkin Myelitis Neuromyelitis Optica Purpura Purpura, Thrombocytopenic Purpura, Thrombocytopenia	6
Romidepsin	Lymphoma, T-Cell, Cutaneous	81
Rucaparib	Carcinoma, Ovarian Epithelial Fallopian Tube Neoplasms Peritoneal Neoplasms Prostatic Neoplasms, Castration- Resistant	74
Ruxolitinib	Graft vs Host Disease Polycythemia Polycythemia Vera Primary Myelofibrosis Thrombocytosis	83
Selinexor	Multiple Myeloma	41
Selumetinib	Neurofibromatosis 1	81
Siltuximab	Giant Lymph Node Hyperplasia	-
Sirolimus	Angiomyolipoma Constriction, Pathologic Coronary Restenosis Eye Diseases Immune System Diseases Kidney Failure, Chronic Lipoma Tuberous Sclerosis	90
Sonidegib	Carcinoma, Basal Cell	-
Sorafenib	Carcinoma, Hepatocellular Carcinoma, Renal Cell Thyroid Neoplasms	94
Sunitinib	Adenoma Carcinoma, Renal Cell Digestive System Neoplasms Gastrointestinal Neoplasms Gastrointestinal Stromal Tumors Intestinal Neoplasms	97
Talazoparib	Breast Neoplasms	27
Tamoxifen	Breast Diseases Cystic Fibrosis Cysts Fibroadenoma Fibrocystic Breast Disease Hemorrhage Menorrhagia Menstruation Disturbances Metrorrhagia Neoplasms	58
Tamsulosin	Calculi Coronary Artery Disease Heart Diseases Hernia Hernia, Inguinal Inflammation Ischemia Lithiasis Lower Urinary Tract Symptoms Myocardial Ischemia Prostatic Hyperplasia Ureteral Calculi Urinary Calculi Urolithiasis Urologic Diseases	-
Temozolomide	Astrocytoma Nervous System Neoplasms	42
Temsirolimus	Carcinoma, Renal Cell	88
Teniposide	Precursor Cell Lymphoblastic Leukemia-Lymphoma	45
Thalidomide	Brain Abscess Immune System Diseases Multiple Myeloma Neoplasms, Plasma Cell	52
Thalidomide Tivozanib		52 87

	Arteritis Neurobenavioral Manifestations Oral Manifestations Psychotic Disorders Schizophrenia Tic Disorders	
Topotecan	Small Cell Lung Carcinoma	40
Toremifene	Breast Neoplasms	11
Trabectedin	Leiomyosarcoma Liposarcoma	-
Trametinib	Carcinoma, Non-Small-Cell Lung Melanoma	85
Trastuzumab	Breast Neoplasms   Neoplasms	71
Tretinoin	Lentigo	68
Triptorelin	Fatty Liver Hypogonadism Infertility, Female Prostatic Neoplasms	67
Tucatinib	Breast Neoplasms	87
Valrubicin	Urinary Bladder Neoplasms	-
Vandetanib	Thyroid Neoplasms	94
Vemurafenib	Melanoma	73
Venetoclax	Leukemia, Lymphocytic, Chronic, B-Cell Leukemia, Myeloid, Acute	-
Vinblastine	Glioma	42
Vincristine	Precursor Cell Lymphoblastic Leukemia-Lymphoma	40
Vinorelbine	Carcinoma, Non-Small-Cell Lung	54
Vismodegib	Carcinoma, Basal Cell	-
Vorinostat	Lymphoma, T-Cell, Cutaneous	80
Zoledronate	Arthritis Bone Marrow Diseases Brain Abscess Chronic Kidney Disease-Mineral and Bone Disorder Chronic Periodontitis HIV Infections Hypersensitivity Infections Kidney Diseases Metabolic Diseases Multiple Myeloma Neoplasms Neoplasms, Plasma Cell Neoplasms, Second Primary Osteitis Osteoarthritis Periodontitis Pleural Effusion, Malignant Prostatic Neoplasms Renal Insufficiency, Chronic Thalassemia Wounds and Injuries	7

# 6. Conclusion

We applied the software package "Genome Enhancer" to a data set that contains *genomics* data. The study is done in the context of *Colorectal Neoplasms*. The data were pre-processed, statistically analyzed and genes carrying sequence variations were identified. Also checked was the enrichment of GO or disease categories among the studied gene sets.

We propose the following drugs as most promising candidates for treating the pathology under study:

Rheumatoid Behavior I Cytokine Release Syndrome I Giant Cell



Erlotinib, seliciclib, Camptothecin and 2,6-Dihydroanthra/1,9-Cd/Pyrazol-6-One

These drugs were selected for acting on the following targets: DYRK1B and HIF1A, which were predicted to be involved in the molecular mechanism of the pathology under study.

The identified molecular mechanism of the studied pathology was predicted to be mainly based on the following key drug targets:



These potential drug targets should be considered as a prospective research initiative for further drug repurposing and drug development purposes. The following drugs were predicted as, matching those drug targets: Tranilast, Bortezomib, Tazarotene, CA4P, Tegafur and uab-30. These drugs should be considered with special caution for research purposes only.

In this study, we came up with a detailed signal transduction network regulating genes carrying sequence variations in the studied pathology. In this network we have revealed the following top master regulators (signaling proteins and their complexes) that play a crucial role in the molecular mechanism of the studied pathology, which can be proposed as the most promising molecular targets for further drug repurposing and drug development initiatives.

- integrins
- PML
- 26S proteasome

Potential drug compounds which can be affecting these targets can be found in the "Finding prospective drug targets" section.

### 7. Methods

# Databases used in the study

Transcription factor binding sites in promoters and enhancers of genes carrying sequence variations were analyzed using known DNA-binding motifs described in the TRANSFAC® library, release 2022.2 (geneXplain GmbH, Wolfenbüttel, Germany) (https://genexplain.com/transfac).

The master regulator search uses the TRANSPATH® database (BIOBASE), release 2022.2 (geneXplain GmbH, Wolfenbüttel, Germany) (https://genexplain.com/transpath). A comprehensive signal transduction network of human cells is built by the software on the basis of reactions annotated in TRANSPATH®.

The information about drugs corresponding to identified drug targets and clinical trials references were extracted from HumanPSD™ database, release 2022.2 (https://genexplain.com/humanpsd).

The Ensembl database release Human104.38 (hg38) (http://www.ensembl.org) was used for gene IDs representation and Gene Ontology (GO) (http://geneontology.org) was used for functional classification of the studied gene set.

### **Genomic data processing**

When analyzing a list of genomic variations (from input vcf file or computed by Genome Enhancer from SNP list or from fastq files), first of all, we compute a specific mutation weight  $(w_1)$  for each variation depending on it's location in gene body and gene flanking regions (-1000 upstream and +1000 downstream of the gene body).

```
w_1 = 0.7 for variations in exon area
```

 $w_1 = 1.3$  for variations in promoter region (-1000bp upstream and 100bp downstream of TSS),

 $w_1 = 1.0$  for variations in other locations.

Next, VCF track (Yes track), provided as input or created by Genome Enhancer from SNP list or fastq files, is compared to Random VCF track (No track) of 10000 random human variations. On both tracks we calculate the score delta values (differences between PWM score values of the TF sites with the reference or with the alternative allele of the considered variation). For each variation we find then the maximal score delta values at each PWM leading either to the gain or to the loss of TF site (with the alternative allele). For selecting the maximum score delta values we consider both directions of DNA strand. Next, by going through all variations we compute two p-values for each PWM – the p-value of site losses and p-value of site gains. The p-values are computed using cumulative Binomial distribution estimating the random chances to observe the found high number of lost or gained TF sites in Yes track in the comparison to the No track. The PWM cut-offs are optimized to obtain the most extreme p-values. We further take top 20 best matrices by p-value from each: gained and lost sites and calculate the mutation weights on the Yes track on the basis of the obtained 40 matrices. Each mutation is assigned with a respective matrix that got the maximum delta value either for the site gain or for the site loss (changed the binding affinity most significantly). This delta is then compared to other delta values that were computed for the respective matrix on the No track. The eventual weight that reflects the transcription factor binding affinity change caused by the mutation is calculated as follows:

```
w_2 = -log10( NoGr / NoAll ), if NoGr > 0

w_2 = -log10( 1.0 / ( 2.0 * NoAll ), if NoGr = 0
```

where NoGr is the number of deltas from the No track that appeared to be greater than the inspected delta and NoAll is the total number of deltas in the No track. The resulting track is then constructed that contains all sites of the initial Yes track together with the additional weights reflecting the transcription factor binding affinity change caused by the mutation.

The list of 40 matrices most affected by variations will be further used in composite modules search described in the next section. Total Gene mutation weight is the sum of the weights  $w_1$  of all variations located inside the gene body and in the gene flanking regions summed up with the weight  $w_2$  that reflects the transcription factor binding affinity change caused by the mutation. This weight is calculated by estimating the importance of a certain mutation in terms of gains or losses of binding sites caused by it. Next, a weighted score is calculated for all genes with the following formula: Weighted score = In\_disease \* In\_transpath \* Gene mutation weight, where

```
In_disease = 2.0 for genes assigned to selected diseases,
In_transpath = 1.5 for genes mapped to Transpath pathways,
and In_disease = In_transpath = 1.0 in all other cases.
```

At the next step, 300 genes with highest weighted score are selected for further CMA model search.

The mutation weights ( $w = w_1 + w_2$ ) are also used to find the regulatory regions of the genes most affected by the variations/SNP. A sliding window of 1100 bp is used to scan through the intronic, 5' and 3' regions of the genes and a region is selected with the highest sum of the mutation weights.

### Methods for the analysis of enriched transcription factor binding sites and composite modules

Transcription factor binding sites in promoters and enhancers of differentially expressed genes were analyzed using known DNA-binding motifs. The motifs are specified using position weight matrices (PWMs) that give weights to each nucleotide in each position of the DNA binding motif for a transcription factor or a group of them.

We search for transcription factor binding sites (TFBS) that are enriched in the enhancers under study as compared to a background set of promoters of housekeeping genes. We denote study and background sets briefly as Yes and No sets. In the current work we used a workflow considering promoter sequences of a standard length of 1100 bp (-1000 to +100). The error rate in this part of the pipeline is controlled by estimating the adjusted p-value (using the Benjamini-Hochberg procedure) in comparison to the TFBS frequency found in randomly selected regions of the human genome (adj.p-value < 0.01).

We have applied the CMA algorithm (Composite Module Analyst) for searching composite modules [7] in the promoters and enhancers of the Yes and No sets. We searched for a composite module consisting of a cluster of 10 TFs in a sliding window of 200-300 bp that statistically significantly separates sequences in the Yes and No sets (minimizing Wilcoxon p-value). Each composite module is forced to include at least one matrix that was identified as matrix causing the significant change in the transcription factor binding affinity as the result of the observed mutation.

# Methods for finding master regulators in networks

We searched for master regulator molecules in signal transduction pathways upstream of the identified transcription factors. The master regulator search uses a comprehensive signal transduction network of human cells. The main algorithm of the master regulator search has been described earlier [3,4]. The goal of the algorithm is to find nodes in the global signal transduction network that may potentially regulate the activity of a set of transcription factors found at the previous step of the analysis. Such nodes are considered as most promising drug targets, since any influence on such a node may switch the transcriptional programs of hundreds

of genes that are regulated by the respective TFs. In our analysis, we have run the algorithm with a maximum radius of 12 steps upstream of each TF in the input set. The error rate of this algorithm is controlled by applying it 10000 times to randomly generated sets of input transcription factors of the same set-size. Z-score and FDR value of ranks are calculated then for each potential master regulator node on the basis of such random runs (see detailed description in [9]). We control the error rate by the FDR threshold 0.05.

# Methods for analysis of pharmaceutical compounds

We seek for the optimal combination of molecular targets (key elements of the regulatory network of the cell) that potentially interact with pharmaceutical compounds from a library of known drugs and biologically active chemical compounds, using information about known drugs from HumanPSD™ and predicting potential drugs using PASS program.

Method for analysis of known pharmaceutical compounds

We selected compounds from HumanPSD™ database that have at least one target. Next, we sort compounds using "Drug rank" that is the sum of the following ranks:

- 1. ranking by "Target activity score" (T-score<sub>PSD</sub>),
- 2. ranking by "Disease activity score" (*D-score<sub>PSD</sub>*),
- 3. ranking by "Clinical validity score".

"Target activity score" ( T-score<sub>PSD</sub>) is calculated as follows:

$$T\text{-}score_{_{PSD}} = -\frac{|T|}{|T| + w(|AT| - |T|))} \sum_{t \in T} log_{10} \left(\frac{rank(t)}{1 + maxRank(T)}\right),$$

where T is set of all targets related to the compound intersected with input list, |T| is number of elements in T, AT and |AT| are set set of all targets related to the compound and number of elements in it, w is weight multiplier, rank(t) is rank of given target, maxRank(T) equals max(rank(t)) for all targets t in T.

We use following formula to calculate "Disease activity score" ( D-score<sub>PSD</sub>):

$$D\text{-}score_{_{P\!S\!D}} = \begin{cases} \sum\limits_{d \in D} \sum\limits_{p \in P} phase(d,p) \\ 0, \ D = \varnothing \end{cases},$$

where D is the set of selected diseases, and if D is empty set, D-score<sub>PSD</sub>=0. P is a set of all known phases for each disease, phase(p,d) equals to the phase number if there are known clinical trials for the selected disease on this phase and zero otherwise. The clinical validity score reflects the number of the highest clinical trials phase (from 1 to 4) on which the drug was ever tested for any pathology.

Method for prediction of pharmaceutical compounds

In this study, the focus was put on compounds with high pharmacological efficiency and low toxicity. For this purpose, comprehensive library of chemical compounds and drugs was subjected to a SAR/QSAR analysis. This library contains 13040 compounds along with their pre-calculated potential pharmacological activities of those substances, their possible side and toxic effects, as well as the possible mechanisms of action. All biological activities are expressed as probability values for a substance to exert this activity (Pa).

We selected compounds that satisfied the following conditions:

- 1. Toxicity below a chosen toxicity threshold (defines as Pa, probability to be active as toxic substance).
- 2. For all predicted pharmacological effects that correspond to a set of user selected disease(s) Pa is greater than a chosen effect threshold
- 3. There are at least 2 targets (corresponding to the predicted activity-mechanisms) with predicted Pa greater than a chosen target threshold.

The maximum Pa value for all toxicities corresponding to the given compound is selected as the "Toxicity score". The maximum Pa value for all activities corresponding to the selected diseases for the given compound is used as the "Disease activity score". "Target activity score" (T-score) is calculated as follows:

$$T\text{-}score(s) = \frac{|T|}{|T| + w(|AT| - |T|)} \sum_{m \in M(s)} \left( pa(m) \sum_{g \in G(m)} IAP(g) optWeight(g) \right),$$

where M(s) is the set of activity-mechanisms for the given structure (which passed the chosen threshold for activity-mechanisms Pa); G(m) is the set of targets (converted to genes) that corresponds to the given activity-mechanism (m) for the given compound; pa(m) is the probability to be active of the activity-mechanism (m), IAP(g) is the invariant accuracy of prediction for gene from G(m); optWeight(g) is the additional weight multiplier for gene. T is set of all targets related to the compound intersected with input list, |T| is number of elements in T, AT and |AT| are set set of all targets related to the compound and number of elements in it, W is weight multiplier.

"Druggability score" (D-score) is calculated as follows: 
$$D\text{-}score(g) = IAP(g) \sum_{s \in S(g)} \sum_{m \in M(s,g)} pa(m),$$

where S(g) is the set of structures for which target list contains given target, M(s,g) is the set of activity-mechanisms (for the given structure) that corresponds to the given gene, pa(m) is the probability to be active of the activity-mechanism (m), IAP(g) is the invariant accuracy of prediction for the given gene.

### 8. References

- 1. Kel A, Voss N, Jauregui R, Kel-Margoulis O, Wingender E. Beyond microarrays: Finding key transcription factors controlling signal transduction pathways. *BMC Bioinformatics.* **2006**;7(S2), S13. doi:10.1186/1471-2105-7-s2-s13
- 2. Stegmaier P, Voss N, Meier T, Kel A, Wingender E, Borlak J. Advanced Computational Biology Methods Identify Molecular Switches for Malignancy in an EGF Mouse Model of Liver Cancer. *PLoS ONE.* **2011**;6(3):e17738. doi:10.1371/journal.pone.0017738
- 3. Koschmann J, Bhar A, Stegmaier P, Kel A, Wingender E. "Upstream Analysis": An Integrated Promoter-Pathway Analysis Approach to Causal Interpretation of Microarray Data. *Microarrays.* **2015**;4(2):270-286. doi:10.3390/microarrays4020270.
- 4. Kel A, Stegmaier P, Valeev T, Koschmann J, Poroikov V, Kel-Margoulis OV, and Wingender E. Multi-omics "upstream analysis" of regulatory genomic regions helps identifying targets against methotrexate resistance of colon cancer. *EuPA Open Proteom.* **2016**;13:1-13. doi:10.1016/j.euprot.2016.09.002
- 5. Michael H, Hogan J, Kel A et al. Building a knowledge base for systems pathology. *Brief Bioinformatics.* **2008**;9(6):518-531. doi:10.1093/bib/bbn038
- Matys V, Kel-Margoulis OV, Fricke E, Liebich I, Land S, Barre-Dirrie A, Reuter I, Chekmenev D, Krull M, Hornischer K, Voss N, Stegmaier P, Lewicki-Potapov B, Saxel H, Kel AE, Wingender E. TRANSFAC and its module TRANSCompel: transcriptional gene regulation in eukaryotes. *Nucleic Acids Res.* 2006;34(90001):D108-D110. doi:10.1093/nar/gkj143
- 7. Kel AE, Gössling E, Reuter I, Cheremushkin E, Kel-Margoulis OV, Wingender E. MATCH: A tool for searching transcription factor binding sites in DNA sequences. *Nucleic Acids Res.* **2003**;31(13):3576-3579. doi:10.1093/nar/gkg585
- 8. Waleev T, Shtokalo D, Konovalova T, Voss N, Cheremushkin E, Stegmaier P, Kel-Margoulis O, Wingender E, Kel A. Composite Module Analyst: identification of transcription factor binding site combinations using genetic algorithm. *Nucleic Acids Res.* **2006**;34(Web Server issue):W541-5.
- 9. Krull M, Pistor S, Voss N, Kel A, Reuter I, Kronenberg D, Michael H, Schwarzer K, Potapov A, Choi C, Kel-Margoulis O, Wingender E. TRANSPATH: an information resource for storing and visualizing signaling pathways and their pathological aberrations. *Nucleic Acids Res.* **2006**;34(90001):D546-D551. doi:10.1093/nar/gkj107
- Boyarskikh U, Pintus S, Mandrik N, Stelmashenko D, Kiselev I, Evshin I, Sharipov R, Stegmaier P, Kolpakov F, Filipenko M, Kel A. Computational master-regulator search reveals mTOR and PI3K pathways responsible for low sensitivity of NCI-H292 and A427 lung cancer cell lines to cytotoxic action of p53 activator Nutlin-3. BMC Med Genomics. 2018;11(1):12. doi:10.1186/1471-2105-7-s2-s13
- 1. Filimonov D, Poroikov V. Probabilistic Approaches in Activity Prediction. Varnek A, Tropsha A. *Cheminformatics Approaches to Virtual Screening*. Cambridge (UK): RSC Publishing. **2008**;:182-216.
- Filimonov DA, Poroikov VV. Prognosis of specters of biological activity of organic molecules. Russian chemical journal. 2006;50(2):66-75 (russ)
- 3. Filimonov D, Poroikov V, Borodina Y, Gloriozova T. Chemical Similarity Assessment Through Multilevel Neighborhoods of Atoms: Definition and Comparison with the Other Descriptors. *ChemInform.* **1999**;39(4):666-670. doi:10.1002/chin.199940210

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In case of any questions please contact us at <a href="mailto:support@genexplain.com">support@genexplain.com</a>

# **Supplementary material**

- 1. Supplementary table 1 Detailed report. Composite modules and master regulators (the most frequently mutated genes in Experiment: short-term survival).
- 2. Supplementary table 2 Detailed report. Pharmaceutical compounds and drug targets.

### **Disclaimer**

Decisions regarding care and treatment of patients should be fully made by attending doctors. The predicted chemical compounds listed in the report are given only for doctor's consideration and they cannot be treated as prescribed medication. It is the physician's responsibility to independently decide whether any, none or all of the predicted compounds can be used solely or in combination for patient treatment purposes, taking into account all applicable information regarding FDA prescribing recommendations for any therapeutic and the patient's condition, including, but not limited to, the patient's and family's medical history, physical examinations, information from various diagnostic tests, and patient preferences in accordance with the current standard of care. Whether or not a particular patient will benefit from a selected therapy is based on many factors and can vary significantly.

The compounds predicted to be active against the identified drug targets in the report are not guaranteed to be active against any particular patient's condition. GeneXplain GmbH does not give any assurances or guarantees regarding the treatment information and conclusions given in the report. There is no guarantee that any third party will provide a refund for any of the treatment decisions made based on these results. None of the listed compounds was checked by Genome Enhancer for adverse side-effects or even toxic effects.

The analysis report contains information about chemical drug compounds, clinical trials and disease biomarkers retrieved from the HumanPSD™ database of gene-disease assignments maintained and exclusively distributed worldwide by geneXplain GmbH. The information contained in this database is collected from scientific literature and public clinical trials resources. It is updated to the best of geneXplain's knowledge however we do not guarantee completeness and reliability of this information leaving the final checkup and consideration of the predicted therapies to the medical doctor.

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