# CDK7 and MNAT1 are promising druggable targets for treating neoplasm metastasis and osteosarcoma that control activity of SMAD2, POU5F1 and TAL1 transcription factors on promoters of differentially expressed genes

Demo User geneXplain GmbH info@genexplain.com Data received on 07/09/2019 ; Run on 19/02/2020 ; Report generated on 19/02/2020

Genome Enhancer release 1.9 (TRANSFAC®, TRANSPATH® and HumanPSD<sup>™</sup> release 2020.1)



#### Abstract

In the present study we applied the software package "Genome Enhancer" to a multiomics data set that contains *transcriptomics and proteomics* data. The study is done in the context of *neoplasm metastasis and osteosarcoma*. The goal of this pipeline is to identify potential drug targets in the molecular network that governs the studied pathological process. In the first step of analysis pipeline discovers transcription factors (TFs) that regulate genes activities in the pathological state. The activities of these TFs are controlled by so-called master regulators, which are identified in the second step of analysis. After a subsequent druggability checkup, the most promising master regulators are chosen as potential drug targets for the analyzed pathology. At the end the pipeline comes up with (a) a list of known drugs and (b) novel biologically active chemical compounds with the potential to interact with selected drug targets.

From the data set analyzed in this study, we found the following TFs to be potentially involved in the regulation of the differentially expressed genes: SMAD2, POU5F1, IRF2, TAL1 and SMAD3. The subsequent network analysis suggested CDK7, LYN, IKBKB, MNAT1 and CCNH as the most promising and druggable molecular targets. Finally, the following drugs were identified as the most promising treatment candidates: Bosutinib, Mesalazine, Flavopiridol, Streptomycin, 1-3 Sugar Ring of Pentamannosyl 6-Phosphate and Uridine.

## 1. Introduction

Recording "-omics" data to measure gene activities, protein expression or metabolic events is becoming a standard approach to characterize the pathological state of an affected organism or tissue. Increasingly, several of these methods are applied in a combined approach leading to large "multiomics" datasets. Still the challenge remains how to reveal the underlying molecular mechanisms that render a given pathological state different from the norm. The disease-causing mechanism can be described by a re-wiring of the cellular regulatory network, for instance as a result of a genetic or epigenetic alterations influencing the activity of relevant genes. Reconstruction of the disease-specific regulatory networks can help identify potential master regulators of the respective pathological process. Knowledge about these master regulators can point to ways how to block a pathological regulatory cascade. Suppression of certain molecular targets as components of these cascades may stop the pathological process and cure the disease.

Conventional approaches of statistical "-omics" data analysis provide only very limited information about the causes of the observed phenomena and therefore contribute little to the understanding of the pathological molecular mechanism. In contrast, the "upstream analysis" method [1-4] applied here has been deviced to provide a casual interpretation of the data obtained for a pathology state. This approach comprises two major steps: (1) analysing promoters and enhancers of differentially expressed genes for the transcription factors (TFs) involved in their regulation and, thus, important for the process under study; (2) re-constructing the signaling pathways that activate these TFs and identifying master regulators at the top of such pathways. For the first step, the database TRANSFAC® [6] is employed together with the TF binding site identification algorithms Match [7] and CMA [8]. The second step involves the signal transduction database TRANSPATH® [9] and special graph search algorithms [10] implemented in the software "Genome Enhancer".

The "upstream analysis" approach has now been extended by a third step that reveals known drugs suitable to inhibit (or activate) the identified molecular targets in the context of the disease under study. This step is performed by using information from HumanPSD<sup>M</sup> database [5]. In addition, new potential small molecular ligands are subsequently predicted for the revealed targets. A general druggability check is performed using a precomputed database of biologcal activities of chemical compounds from a library of about 13000 pharmaceutically most active compounds. The spectra of biological activities are computed using the program PASS on the basis of a (Q)SAR approach [11-13].

## 2. Data

For this study the following experimental data was used:

Table 1. Experimental datasets used in the study

File name	Data type
Proteomics	Proteomics
RNAseq	Transcriptomics



Figure 1. Annotation diagram of experimental data used in this study. With the colored boxes we show those sub-categories of the data that are compared in our analysis.

## 3. Results

We have compared the following conditions: Myc\_induce versus Control.

## 3.1. Identification of target genes

In the first step of the analysis **target genes** were identified from the uploaded experimental data. We applied the Limma tool (R/Bioconductor package integrated into our pipeline) and compared gene expression in the following sets: "Myc\_induce" with "Control". Limma calculated the LogFC (the logarithm to the base 2 of the fold change between different conditions), the p-value and the adjusted p-value (corrected for multiple testing) of the observed fold change. As a result, we detected 5047 upregulated genes (LogFC>0) out of which 1195 genes were found as significantly upregulated (p-value<0.1) and 4524 downregulated genes (LogFC<0) out of which 1169 genes were significantly downregulated (p-value<0.1). See tables below for the top significantly up- and downregulated genes. Below we call **target genes** the full list of up- and downregulated genes revealed in our analysis (see tables in Supplementary section).

Table 2. Top ten significant **up-regulated** genes in Myc\_induce vs. Control. **See full table**  $\rightarrow$ 

ID	Gene symbol	Gene description	logFC	P.Value	adj.P.Val
ENSG00000136997	MYC	v-myc avian myelocytomatosis viral oncogene homolog	5.96	7.45E-6	7.13E-2
ENSG00000164076	CAMKV	CaM kinase like vesicle associated	4.08	8.1E-5	0.13
ENSG00000120738	EGR1	early growth response 1	3.51	5.46E-4	0.14
ENSG00000173110	HSPA6	heat shock protein family A (Hsp70) member 6	3.14	1.66E-4	0.13
ENSG00000123360	PDE1B	phosphodiesterase 1B	2.85	1.08E-4	0.13
ENSG00000137571	SLCO5A1	solute carrier organic anion transporter family member 5A1	2.79	9.53E-5	0.13
ENSG0000078549	ADCYAP1R1	ADCYAP receptor type I	2.69	2.44E-3	0.14
ENSG00000143333	RGS16	regulator of G-protein signaling 16	2.69	2.47E-4	0.13
ENSG00000170345	FOS	Fos proto-oncogene, AP-1 transcription factor subunit	2.57	4.12E-3	0.15
ENSG00000117322	CR2	complement C3d receptor 2	2.46	2.57E-4	0.13

Table 3. Top ten significant **down-regulated** genes in Myc\_induce vs. Control.

See full table  $\rightarrow$ 

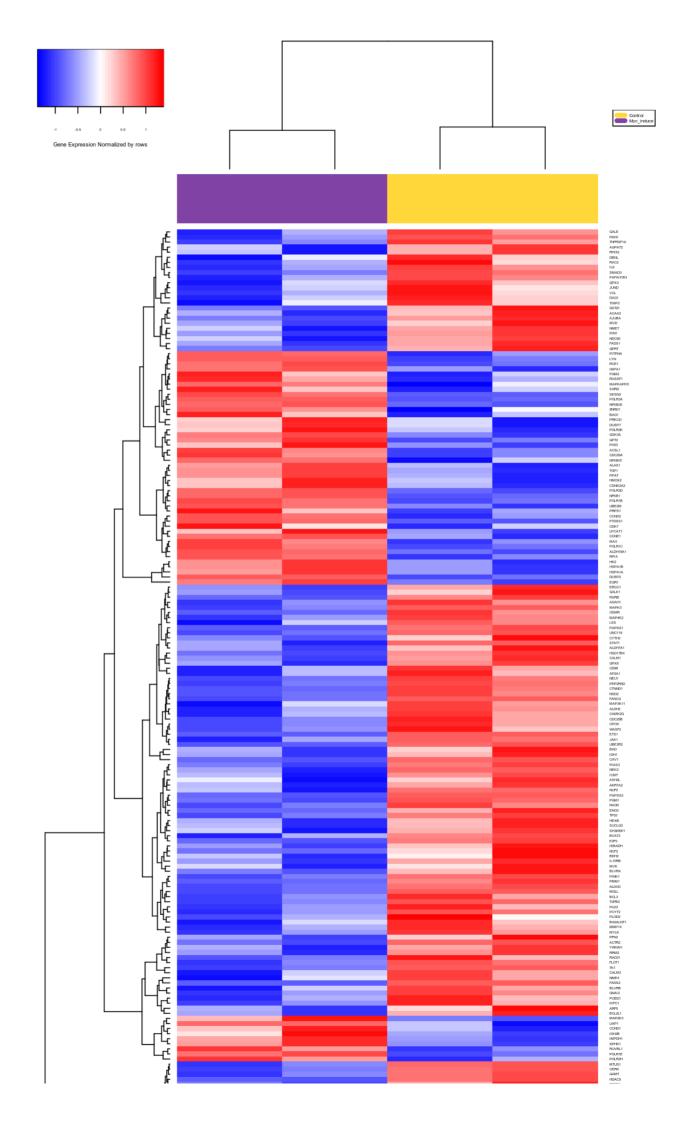
ID	Gene symbol	Gene description	logFC	P.Value	adj.P.Val
ENSG00000116774	OLFML3	olfactomedin like 3	-3.06	1.11E-4	0.13
ENSG00000138131	LOXL4	lysyl oxidase like 4	-2.62	8.88E-4	0.14
ENSG0000187867	PALM3	paralemmin 3	-2.62	2.65E-3	0.14
ENSG00000205542	TMSB4X	thymosin beta 4, X-linked	-2.58	2.22E-4	0.13
ENSG00000158825	CDA	cytidine deaminase	-2.54	3.49E-4	0.13
ENSG00000127129	EDN2	endothelin 2	-2.49	3.28E-4	0.13
ENSG00000182667	NTM	neurotrimin	-2.48	4.08E-4	0.13
ENSG00000114115	RBP1	retinol binding protein 1	-2.46	1.06E-4	0.13
ENSG00000132746	ALDH3B2	aldehyde dehydrogenase 3 family member B2	-2.35	1.93E-4	0.13
ENSG00000188042	ARL4C	ADP ribosylation factor like GTPase 4C	-2.29	1.87E-3	0.14

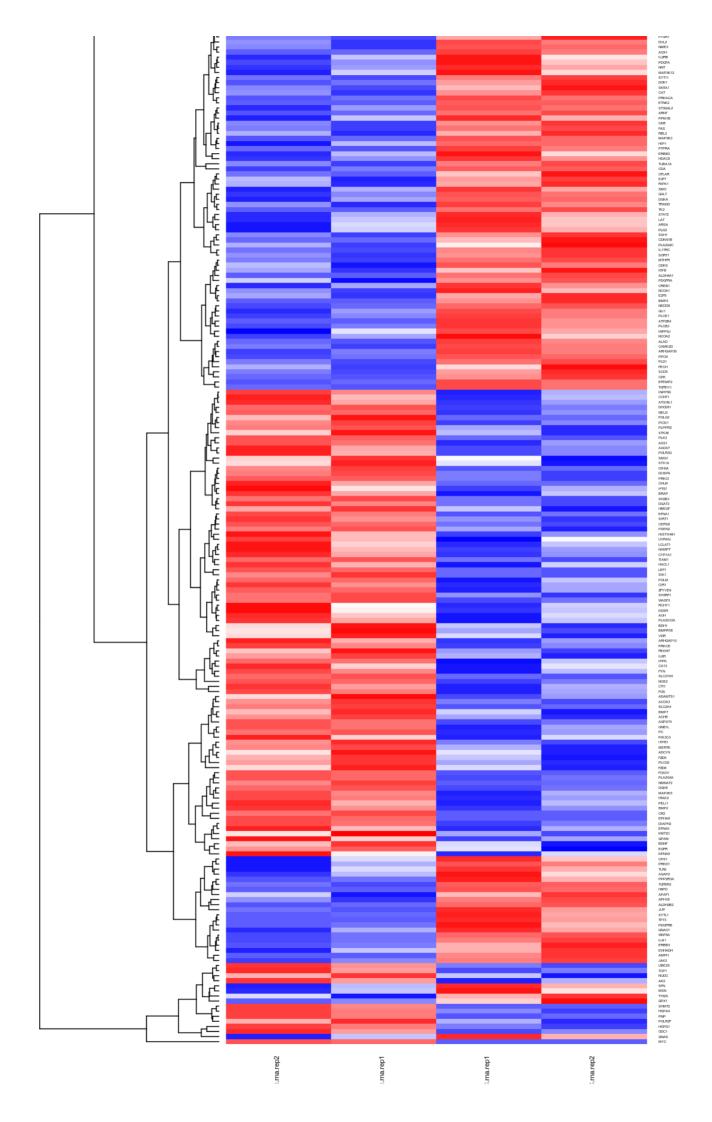
#### 3.2. Functional classification of genes

A functional analysis of differentially expressed genes was done by mapping the significant up-regulated and significant down-regulated genes to several known ontologies, such as Gene Ontology (GO), disease ontology (based on HumanPSD<sup>™</sup> database) and the ontology of signal transduction and metabolic pathways from the TRANSPATH® database. Statistical significance was computed using a binomial test. Figures 3-8 show the most significant categories.

#### Heatmap of differentially expressed genes in Myc\_induce vs. Control

A heatmap of all differentially expressed genes playing a potential regulatory role in the system (enriched in TRANSPATH® pathways) is presented in Figure 2.





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Figure 2. Heatmap of genes enriched in Transpath categories. The colored bar at the top shows the types of the samples according to the legend in the upper right corner. See full diagram  $\rightarrow$ 

## Up-regulated genes in Myc\_induce vs. Control:

1195 significant up-regulated genes were taken for the mapping.

Myc

#### GO (biological process)

				bio	ological_proc	ess Gene On	tology treem	ар					
RNA metaboli	c process	nucleic acid metabolic process	s macromolecule biosynthetic process	regulation of RNA metabolic process	regulation of cellular biosynthetic process	regulation of cellular macromolecule biosynthetic process	RNA modification	IRNA	RNA methylation	RNA proc	Cessing (BAA 3-eed	cellula biosynth proces	etic process
				regulation of biosynthetic process	regulation of transcription, DNA-templated	regulation of macromolecule biosynthetic process	tRNA modification	rRNA	ation R	NA pr	ocessing	orgar biosyn	nic substance
cellular nitrogen compound biosynthetic process	nucleic acid-templa transcripti	ated process	nucleobase-containing compound biosynthetic process	_	tion of c romole		RNA phosphodiester bond hydrolysis	1994A muchik phosphodester phospho band bar hydrolysis, hydro endonucleolytic	acid denter d hysta	r nitrogen d metabolic ocess	nitrogen con metabolic p	npound rocess	cellular macromolecule metabolic proces
transcription, DNA-templated	cellular macromole biosynthe	cule	cess cyclic compound biosynthetic	biosyn nucleobase-con compound met process	~	OCESS eterocycle ibolic process	involved in rRNA processing RNA phos	enderski enderski softerfor (152)-1790 ALSD-1790 process phodieste /drolysis	cellular compoun	nitrogen d metabolio ocess	c nitrogen cor metabolic p		cellular macromolecul netabolic proce
nucleic a	process	aromatic compo biosynthetic pro	cess				organic cycli metabolio	c compound	180.04	ary polic	metabolic proc	cess trans elor from poly	scription ngation mRNA m <b>transcription</b> m <b>transcription</b>
ncRNA metaboli	c process	rRNA processing r	RNA metabolic process	cellular aromatic cellular aromati	c compound me		organic cycli metabolic mitochondrial RNA metabolic process		metal proc matrix metabolic	ess plecule process	transcription initiation from RNA polymerase transcription initiation from RNA polymerase regulation	termination RNA polymr I transcri termination of RN polymer	lymerase I promo on of regulatio herase of gene ofion expression (A regulation
ncRNA proce				ribonucleo complex bio		cellular component biogenesis	mitochon metabolic cellular metal	c process	metabolic	plecule process	of nitrogen compound metabolic process	regulation macromole metabol proces	of cellul cule metabo
ncR	NA me	tRNA processing transport	rf94A II promoter EU-rf94A SU-rf94A snf94A rf1NA transcription	ribosome bio	la I	ribosomal Irge subunit biogenesis x biogenesis	cellular metal	polic proces	organic su	Ibstance	ranscription from RNA polymerase III promoter	regulation of metabolic process	regulation process

Figure 3. Enriched GO (biological process) of up-regulated genes in Myc\_induce vs. Control. Full classification  $\rightarrow$ 

TRANSPATH® Pathways (2020.1)

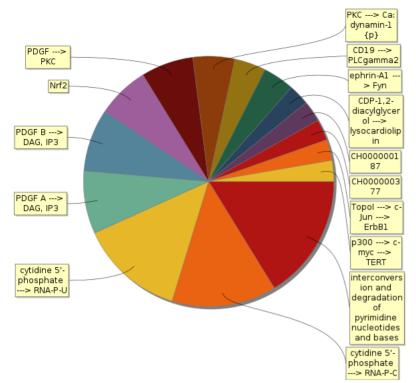
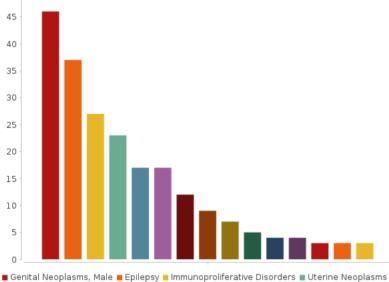


Figure 4. Enriched TRANSPATH® Pathways (2020.1) of up-regulated genes in Myc\_induce vs. Control. Full classification  $\rightarrow$ 

## HumanPSD(TM) disease (2020.1)



🔳 Uterine Cervical Diseases 🔳 Uterine Cervical Neoplasms 🔳 Wilms Tumor

🔳 Triple Negative Breast Neoplasms 🔳 Eye Abnormalities 🔳 Epilepsies, Partial 🔳 Coloboma

🔳 Epilepsy, Temporal Lobe 📕 Aniridia 📕 Iris Diseases 📕 Mandibulofacial Dysostosis

Figure 5. Enriched HumanPSD(TM) disease (2020.1) of up-regulated genes in Myc\_induce vs. Control. The size of the bars correspond to the number of biomarkers of the given disease found among the input set. Full classification  $\rightarrow$ 

## Down-regulated genes in Myc\_induce vs. Control:

1169 significant down-regulated genes were taken for the mapping.

#### GO (biological process)

regulation of cell motility	regulation locomotion		negative regulation of cellular component	single-organism li catabolic process	pid catabolic process	catabolic process	cofactor metabolic proce	ess me	enzyme etabolic rocess	cellular compone organization	nt actin filament-based process	cell death
regulation of cellular component movement	negative regulation of locomotion	positive regulation of cell migration	on regulation of cell		-	carboxylic acid catabolic process fatty acid ta-oxidation	heme metabolic process oxidoreduction coenzyme cofactor meta	tetrapostvinagen IX metabolic process tetrap		cellular compone organization o biogenesis cellular compor organizatior	actin filament-based	cell death process
regulation of cell migration	negative regulation of cell motility positive	of cellu	ion regulation lar of ent locomotion	catabolic process single-organisn response to hypoxia		response	process mitotic sister chromatid segregation	chron		regulation of negative upramolecular fiber organization organization	n of cular	negative regulation of cellular process
-	negative reg	r 🗕 🚽	ility f negative	response to		abiotic stimulus	sister chromati segrecation mitotic chromatid s	sister	ation	regulation of supramolecula fiber organization		negative regulation of biological process negative regulation of biological process
	1	oskeletor janizatior	organization	decreased oxygen levels	response to hyperoxia <b>to hyp</b>	increased	organelle organization	single-or orgar organiz	nelle	regulation of actin filament-based proce regulation of actin	cytoskeleton organization	positive regulation of biological process
	sitive regulation ellular component organization regulation of	microb	ubule regulation of cation or organelle	regulation of signa transduction	-	ulation of	single-or organelle or single-organ	rganiza	ation	ytoskeleton organiza regulation of actin filament-based proce single-organism	regulation of	
regulation of cy negative ne regulation regu	icrotubule-base /toSkeletor agative r ulation of re- operation	n organ negative egulation of protein	barbed-end actin	regulation of cell communication regulation of intrace positive regulation	intra signal t Ilular signal ti	ulation of acellular ransduction ransduction a regulation	single-organi	lem nro		cellular process		single-organism metabolic process response to stress morphogene
component deve organization ne reg of j	egative n julation reg protein prote	lymerization negative gulation of ein comple: sassembly	filament capping actin filament <sup>X</sup> capping	positive regulation response to stimult	is of s	ignaling regulation of	regulation developmen process	of	regulation of cell fferentiation	cellular proces regulation of cellula component organizat	on of cellular process on oxidation-reduction	esponse muscular septum o stress morphogen
regulation of cell projection	e regulation re	egulation of protein olymerization	regulation of protein polymerization	signal transduction		nmunication	regulat developmen		0055	regulation of cellula component organizat		membranous septum membrahous septur morphogenesis

Figure 6. Enriched GO (biological process) of down-regulated genes in Myc\_induce vs. Control. Full classification  $\rightarrow$ 

## TRANSPATH® Pathways (2020.1)

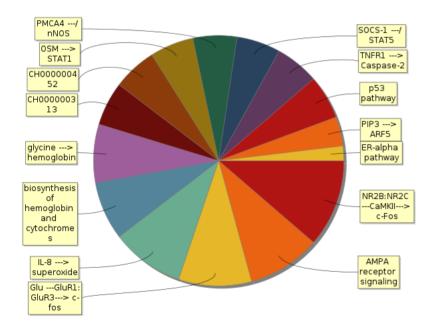
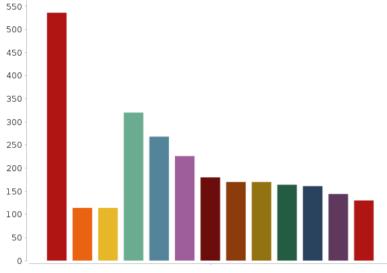


Figure 7. Enriched TRANSPATH® Pathways (2020.1) of down-regulated genes in Myc\_induce vs. Control. Full classification  $\rightarrow$ 

HumanPSD(TM) disease (2020.1)



Digestive System Neoplasms

Neoplasms by Site Endocrine Gland Neoplasms Neoplasms by Histologic Type

📕 Adnexal Diseases 📕 Ovarian Diseases 🔳 Skin and Connective Tissue Diseases

🔳 Ovarian Neoplasms 🔳 Skin Diseases 📕 Neoplasms, Glandular and Epithelial

Figure 8. Enriched HumanPSD(TM) disease (2020.1) of down-regulated genes in Myc\_induce vs. Control. The size of the bars correspond to the number of bio-markers of the given disease found among the input set. Full classification  $\rightarrow$ 

## 3.3. Identification of proteins

In the first step of the proteome data analysis target proteins were identified from the uploaded experimental data (the list of 4665 proteins) and were converted to corresponding genes. These genes were used in the further steps of analysis.

Table 4. Top ten the list of genes provided as input in Myc\_induce. See full table

See full table $\rightarrow$			
ID	Gene description	Gene symbol	Proteomics_avr
ENSG00000173598	nudix hydrolase 4	NUDT4	4.36
ENSG00000100335	mitochondrial elongation factor 1	MIEF1	3.8
ENSG00000115884	syndecan 1	SDC1	3.62
ENSG00000102910	lon peptidase 2, peroxisomal	LONP2	3.3
ENSG00000179046	tripartite motif family like 2	TRIML2	2.87
ENSG00000114648	kelch like family member 18	KLHL18	2.76
ENSG00000170525	6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 3	PFKFB3	2.69
ENSG00000120949	TNF receptor superfamily member 8	TNFRSF8	2.46
ENSG00000188158	NHS actin remodeling regulator	NHS	2.46
ENSG00000119599	DDB1 and CUL4 associated factor 4	DCAF4	2.42

#### 3.4. Functional classification of expressed proteins

A functional analysis of expressed proteins was done by mapping the protein IDs to several known ontologies, such as Gene Ontology (GO), disease ontology (based on HumanPSD™ database) and the ontology of signal transduction and metabolic pathways from the TRANSPATH® database. Statistical significance was computed using a binomial test. Figures 9-11 show the most significant categories.

## The list of proteins provided as input in Myc\_induce:

4653 the list of genes provided as input genes were taken for the mapping.

#### GO (biological process)

					biological_p	rocess Gene	e Ontology tr	eemap				
cellular localization	intracell transpo		establishment o localization in cell	nucleobase-containi compound metabol process		cellular macromolecule catabolic process	macromolecul catabolic process	le organic substance catabolic process	mitotic cell cycle process	inuclear fiss	cycle amide	peptide biosynthetic process
macromolecule localization	protein localization	intrace prote trans	ein substance	cellular aromatic compound metabolic	nucleic acid metabolic process	cellular catabolic process cellular ma	matrician departer matrician catalate process cromolecule cat	protection of the protein protection of the protection of the protection protection protection of the	cell cycle process		ase sition ord ord ord ord ord ord process process process process process process process	metabolic proceas organonitrogen compound biosynthetic
transport	stablishment of protein localization Ilar prot	intrac tran single-c	organism cellular sport organism ular ransport			cellular comp organizatio biogenes	n or comp	bonent of p hization local to org	ishment rotein ization ganelle blishment of pro-	gene expres regulation gene expres regulation geting raile posttrar	of of translation organiza	
mRNA metabo process	cata pro nucleobas compoun	RNA abolic ocess e-containing el catabolic ocess	RNA catabolic process nucker-transcribed mPINA catabolic process, nonsense-mediated decay	cellular component assembly	protein complex subunit organization	cellular con symbiosis, encompassin mutualism thro	g cellular	loca	lization to orga cellular abolic process	nelle gene e		ganization selection cellular macromolecule netabolic process
	compoun	nitrogen id catabolic icess	aromatic compound catabolic process organic cyclic compound catabolic	cellular macromolecular complex assemb cellular macromole		parasitism viral proces <b>vira</b>				metabolic proces	nitrogen compound metabolic process organic cyclic compound n	cellular macromolecule metabolic process cellular protein metabolia process <b>protein</b>
mRNA r		A r	rRNA metabolic process	RNA processin	g RNA splicing	nuclear transport nucleocytoplasmi	export	RNA export from nucleus	process	organic substance metabolic	compound	netabolic process
ncRNA processi	tRNA ing metabol	ic brocessi	amino ndRNA acid transorption		RNA splicing, via transesterification reactions	transport <b>nucle</b> a protein localiz to organel		port biosy	nic substance mthetic process lular nitrogen ound metabolic process	process primary metabolic process	macromolecule metabolic process macromolecule metabolic process	DNA metabolic process
ncRNA	snRNA metabolic metabolic metabolic	aminoad for pro	VA srf94A srf94A ranscriptionTranscriptio from 194A obtin obtin aBOn II sorondar	transesterification reactions with	NA splicing, RNA 3'-end via processing liceosome rocess processing	cellular macromoleo	localizatio	reticulum cel	lular nitrogen bound metabolic process	primary metabolic process	cellular process	nitrogen compound biosynthetic

Figure 9. Enriched GO (biological process) of the list of proteins provided as input in  $Myc_induce$ . Full classification  $\rightarrow$ 

## TRANSPATH® Pathways (2020.1)

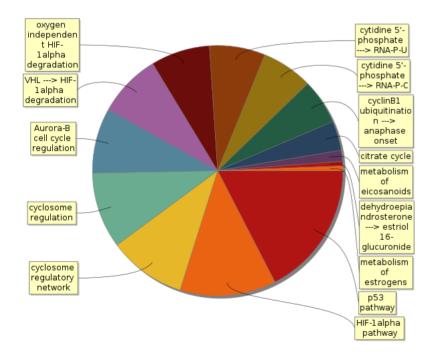
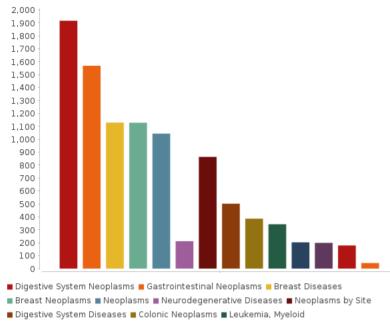


Figure 10. Enriched TRANSPATH® Pathways (2020.1) of the list of proteins provided as input in Myc\_induce. Full classification  $\rightarrow$ 

HumanPSD(TM) disease (2020.1)



🔳 Neoplasms, Adipose Tissue 🔳 Liposarcoma 📕 Liposarcoma, Myxoid 📕 Mitochondrial Diseases

Figure 11. Enriched HumanPSD(TM) disease (2020.1) of the list of proteins provided as input in Myc\_induce. The size of the bars correspond to the number of bio-markers of the given disease found among the input set.

#### Full classification $\rightarrow$

## 3.5. Comparison plot of transcriptome and proteome

After the analysis of transcriptome and proteome data they were compared with each other. Below we plot 9578 genes and 4653 proteins.

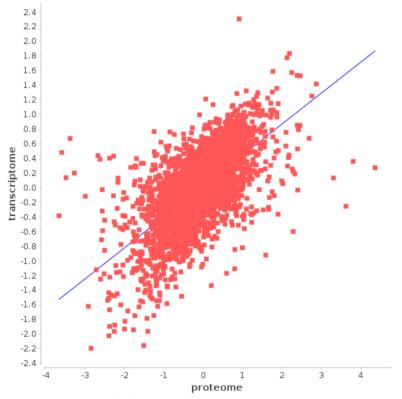


Figure 12. Comparison plot of comparison proteome vs transcriptome. X axis: protein expression value - Proteomics\_avr. Y axis: LogFC of differential gene expression. Full comparison  $\rightarrow$ 

Comparison of up-regulated genes (transcriptome data) and the list of proteins provided as input (proteome data)

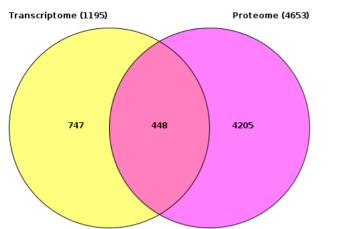


Figure 13. Intersection of up-regulated genes and the list of proteins provided as input See full diagram  $\rightarrow$ 

#### Comparison of down-regulated genes (transcriptome data) and the list of proteins provided as input (proteome data)

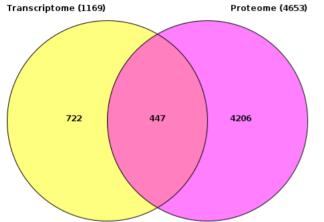


Figure 14. Intersection of down-regulated genes and the list of proteins provided as input See full diagram  $\rightarrow$ 

## 3.6. Analysis of enriched transcription factor binding sites and composite modules

In the next step a search for transcription factors binding sites (TFBS) was performed in the regulatory regions of the **target genes** by using the TF binding motif library of the **TRANSFAC®** database. We searched for so called **composite-modules** that act as potential condition-specific **enhancers** of the **target genes** in their upstream regulatory regions (-1000 bp upstream of transcription start site (TSS)) and identify transcription factors regulating activity of the genes through such **enhancers**.

Classically, **enhancers** are defined as regions in the genome that increase transcription of one or several genes when inserted in either orientation at various distances upstream or downstream of the gene [8]. Enhancers typically have a length of several hundreds of nucleotides and are bound by multiple transcription factors in a cooperative manner [9].

We applied the Composite Module Analyst (CMA) [8] method to detect such potential enhancers, as targets of multiple TFs bound in a cooperative manner to the regulatory regions of the genes of interest. CMA applies a genetic algorithm to construct a generalized model of the enhancers by specifying combinations of TF motifs (from TRANSFAC®) whose sites are most frequently clustered together in the regulatory regions of the studied genes. CMA identifies the transcription factors that through their cooperation provide a synergistic effect and thus have a great influence on the gene regulation process.

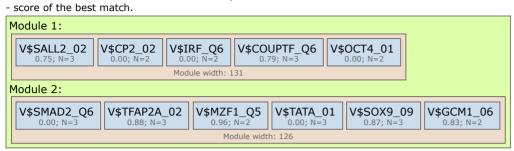
#### Enhancer model potentially involved in regulation of target genes (up-regulated genes in Myc\_induce vs. Control).

To build the most specific composite modules we choose genes as the input of CMA algorithm. The obtained CMA model is then applied to compute CMA score for all up-regulated genes.

The model consists of 2 module(s). Below, for each module the following information is shown:

- PWMs producing matches,

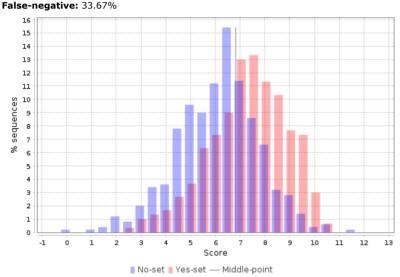
- number of individual matches for each PWM,



Model score (-p\*log10(pval)): 10.03 Wilcoxon p-value (pval): 2.52e-21 Penalty (p): 0.487 Average yes-set score: 7.27 Average no-set score: 6.12

AUC: 0.70 Middle-point: 6.82

False-positive: 33.20%



#### See model visualization table $\rightarrow$

Table 5. List of top ten up-regulated genes in Myc\_induce vs. Control with identified enhancers in their regulatory regions. **CMA score** - the score of the CMA model of the enhancer identified in the regulatory region. **See full table**  $\rightarrow$ 

Ensembl IDs	Gene symbol	Gene description	CMA score	Factor names
ENSG00000154122	ANKH	ANKH inorganic pyrophosphate transport regulator	14.8	SALL2(h), COUP-TF1(h),COUP-TF2(h), MZF-1(h), GCMa(h), CP2(h), Smad2(h), Oct3(h)
ENSG0000083857	FAT1	FAT atypical cadherin 1	14.74	CP2(h), COUP-TF1(h),COUP-TF2(h), Oct3(h), IRF-1(h),IRF-2(h),IRF-3(h),IRF-4(h),IRF-5(h),IRF-6(h),IRF-7(h),IRF-8(h), Smad2(h), MZF-1(h), Sox-9(h)
ENSG00000120071	KANSL1	KAT8 regulatory NSL complex subunit 1	14.67	GCMa(h), MZF-1(h), Smad2(h), COUP-TF1(h),COUP-TF2(h), TBP(h), Oct3(h), Sox- 9(h)
ENSG00000105197	TIMM50	translocase of inner mitochondrial membrane 50	14.5	Sox-9(h), COUP-TF1(h),COUP-TF2(h), MZF-1(h), Smad2(h), SALL2(h), TBP(h), GCMa(h)
ENSG00000107951	MTPAP	mitochondrial poly(A) polymerase	14.41	Sox-9(h), MZF-1(h), TBP(h), Smad2(h), CP2(h), COUP-TF1(h),COUP-TF2(h), Oct3(h)
ENSG00000119950	MXI1	MAX interactor 1, dimerization protein	14.39	COUP-TF1(h),COUP-TF2(h), Smad2(h), Sox-9(h), IRF-1(h),IRF-2(h),IRF-3(h),IRF-4(h),IRF-5(h),IRF-6(h),IRF-7(h),IRF-8(h), TBP(h), MZF-1(h), Oct3(h)
ENSG00000168807	SNTB2	syntrophin beta 2	14.33	TBP(h), Smad2(h), Oct3(h), Sox-9(h), IRF-1(h),IRF-2(h),IRF-3(h),IRF-4(h),IRF- 5(h),IRF-6(h),IRF-7(h),IRF-8(h), COUP-TF1(h),COUP-TF2(h), CP2(h)
ENSG0000138386	NAB1	NGFI-A binding protein 1	14.31	TBP(h), Smad2(h), Sox-9(h), GCMa(h), CP2(h), IRF-1(h),IRF-2(h),IRF-3(h),IRF- 4(h),IRF-5(h),IRF-6(h),IRF-7(h),IRF-8(h), Oct3(h)
ENSG00000144228	SPOPL	speckle type BTB/POZ protein like	14.27	COUP-TF1(h),COUP-TF2(h), Smad2(h), Sox-9(h), MZF-1(h), TBP(h), IRF-1(h),IRF-2(h),IRF-3(h),IRF-4(h),IRF-5(h),IRF-6(h),IRF-7(h),IRF-8(h), GCMa(h)
ENSG0000070444	MNT	MAX network transcriptional repressor	14.25	SALL2(h), Sox-9(h), Smad2(h), COUP-TF1(h),COUP-TF2(h), CP2(h), MZF-1(h), IRF-1(h),IRF-2(h),IRF-3(h),IRF-4(h),IRF-5(h),IRF-6(h),IRF-7(h),IRF-8(h)

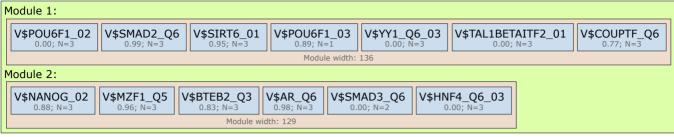
#### Enhancer model potentially involved in regulation of target genes (down-regulated genes in Myc\_induce vs. Control).

To build the most specific composite modules we choose genes as the input of CMA algorithm. The obtained CMA model is then applied to compute CMA score for all down-regulated genes.

The model consists of 2 module(s). Below, for each module the following information is shown:

- PWMs producing matches,

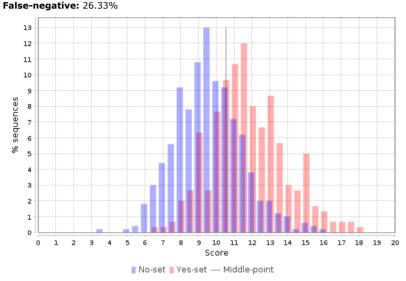
- number of individual matches for each PWM, - score of the best match.



Model score (-p\*log10(pval)): 19.09 Wilcoxon p-value (pval): 6.24e-42 Penalty (p): 0.463 Average yes-set score: 11.76 Average no-set score: 9.59

AUC: 0.79 Middle-point: 10.54

False-positive: 28.00%



#### See model visualization table $\rightarrow$

Table 6. List of top ten down-regulated genes in Myc\_induce vs. Control with identified enhancers in their regulatory regions. **CMA score** - the score of the CMA model of the enhancer identified in the regulatory region. **See full table**  $\rightarrow$ 

Ensembl IDs	Gene symbol	Gene description	CMA score	Factor names
ENSG00000101997	CCDC22	coiled-coil domain containing 22	20.18	AR(h), BTEB2(h), MZF-1(h), Smad3(h), HNF-4alpha(h), SIR2L6(h), COUP- TF1(h),COUP-TF2(h)
ENSG0000063601	MTMR1	myotubularin related protein 1	19.05	Smad2(h), COUP-TF1(h),COUP-TF2(h), YY1(h), POU6F1(h), ITF-2(h),Tal- 1(h), Smad3(h), HNF-4alpha(h)
ENSG00000105048	TNNT1	troponin T1, slow skeletal type	18.88	AR(h), COUP-TF1(h),COUP-TF2(h), HNF-4alpha(h), ITF-2(h),Tal-1(h), POU6F1(h), YY1(h), SIR2L6(h)
ENSG00000148337	CIZ1	CDKN1A interacting zinc finger protein 1	18.82	BTEB2(h), MZF-1(h), nanog(h), HNF-4alpha(h), Smad3(h), SIR2L6(h), ITF- 2(h),Tal-1(h)
ENSG00000122861	PLAU	plasminogen activator, urokinase	18.82	COUP-TF1(h),COUP-TF2(h), Smad2(h), POU6F1(h), ITF-2(h),Tal-1(h), SIR2L6(h), YY1(h), nanog(h)
ENSG00000115461	IGFBP5	insulin like growth factor binding protein 5	18.72	AR(h), Smad3(h), BTEB2(h), MZF-1(h), HNF-4alpha(h), YY1(h), COUP- TF1(h),COUP-TF2(h)
ENSG0000092051	JPH4	junctophilin 4	18.53	Smad3(h), BTEB2(h), MZF-1(h), ITF-2(h),Tal-1(h), nanog(h), AR(h), POU6F1(h)
ENSG00000113721	PDGFRB	platelet derived growth factor receptor beta	18.31	Smad3(h), HNF-4alpha(h), MZF-1(h), BTEB2(h), nanog(h), ITF-2(h),Tal- 1(h), COUP-TF1(h),COUP-TF2(h)
ENSG00000169045	HNRNPH1	heterogeneous nuclear ribonucleoprotein H1	18.18	AR(h), MZF-1(h), BTEB2(h), HNF-4alpha(h), ITF-2(h),Tal-1(h), Smad3(h), YY1(h)
ENSG00000130176	CNN1	calponin 1	18.03	Smad2(h), YY1(h), COUP-TF1(h),COUP-TF2(h), ITF-2(h),Tal-1(h), SIR2L6(h), POU6F1(h), nanog(h)

On the basis of the enhancer models we identified the following transcription factors potentially regulating the *target genes* of our interest. We found 18 and 14 transcription factors controlling expression of up- and down-regulated genes respectively (see Tables 7-8).

Table 7. Transcription factors of the predicted enhancer model potentially regulating the differentially expressed genes (up-regulated genes in Myc\_induce vs. Control). **Yes-No ratio** is the ratio between frequencies of the sites in Yes sequences versus No sequences. It describes the level of the enrichment of binding sites for the indicated TFin the regulatory target regions. **Regulatory score** is the measure of involvement of the given TF in the controlling of expression of genes that encode master-regulators presented below (through positive feedback loops). **See full table**  $\rightarrow$ 

ID	Gene symbol	Gene description	Regulatory score	Yes-No ratio
MO000057829	SMAD2	SMAD family member 2	5.1	1.21
MO000056618	POU5F1	POU class 5 homeobox 1	4.32	1.74
MO000007691	IRF2	interferon regulatory factor 2	4.29	1.3
MO000024736	NR2F1	nuclear receptor subfamily 2 group F member 1	4.02	8.33
MO000285816	IRF3	interferon regulatory factor 3	3.93	1.3
MO000117988	TFCP2	transcription factor CP2	3.9	1.34
MO000007703	IRF7	interferon regulatory factor 7	3.88	1.3
MO000007686	IRF1	interferon regulatory factor 1	3.75	1.3
MO000021896	ТВР	TATA-box binding protein	3.64	1.26
MO000018993	SOX9	SRY-box 9	3.61	1.63

Table 8. Transcription factors of the predicted enhancer model potentially regulating the differentially expressed genes (down-regulated genes in Myc\_induce vs. Control). **Yes-No ratio** is the ratio between frequencies of the sites in Yes sequences versus No sequences. It describes the level of the enrichment of binding sites for the indicated TFin the regulatory target regions. **Regulatory score** is the measure of involvement of the given TF in the controlling of expression of genes that encode master-regulators presented below (through positive feedback loops). **See full table**  $\rightarrow$ 

ID	Gene symbol	Gene description	Regulatory score	Yes-No ratio
MO000032489	TAL1	TAL bHLH transcription factor 1, erythroid differentiation factor	5.14	1.34
MO000057832	SMAD3	SMAD family member 3	5.08	1.69
MO000057829	SMAD2	SMAD family member 2	4.83	1.66
MO000021454	AR	androgen receptor	4.54	11.68
MO000078913	YY1	YY1 transcription factor	4.14	1.16
MO000134485	NANOG	Nanog homeobox	4.1	1.33
MO000027755	HNF4A	hepatocyte nuclear factor 4 alpha	3.87	1.56
MO000028320	POU6F1	POU class 6 homeobox 1	3.78	1.39
MO000142283	SIRT6	sirtuin 6	3.59	1.2
MO000026229	KLF5	Kruppel like factor 5	3.5	1.3

## 3.7. Finding master regulators in networks

In the second step of the upstream analysis common regulators of the revealed TFs were identified. Using proteomics data we selected differentially expressed proteins that are involved in signal transduction pathways and used these proteins as "context set" [4] in the algorithm of identification of master regulators. These master regulators appear to be the key candidates for therapeutic targets as they have a master effect on regulation of intracellular pathways that activate the pathological process of our study. The identified master regulators are shown in Tables 9-10.

Table 9. Master regulators that may govern the regulation of **up-regulated** genes in Myc\_induce vs. Control. **Total rank** is the sum of the ranks of the master molecules sorted by keynode score, CMA score, transcriptomics and proteomics data. **See full table**  $\rightarrow$ 

ID	Master molecule name	Gene symbol	Gene description	Contained in proteome set	Total rank	logFC (transcriptome)
MO000031189	PKCdelta(h)	PRKCD	protein kinase C delta	1	100	0.86
MO000059577	PKCdelta(h)	PRKCD	protein kinase C delta	1	132	0.86
MO000056654	p300(h)	EP300	E1A binding protein p300	1	134	0.26
MO000022058	Lyn(h)	LYN	LYN proto-oncogene, Src family tyrosine kinase	1	147	0.46
MO000117508	TC-PTP(h)	PTPN2	protein tyrosine phosphatase, non-receptor type 2	1	149	0.33
MO000021902	TFIIH-CAK(h)	CCNH, CDK7, MNAT1	MNAT1, CDK activating kinase assembly factor, cyclin H, cyclin dependent kinase 7	1	152	0.63
MO00009386	MEK1(h)	MAP2K1	mitogen-activated protein kinase kinase 1	1	176	0.22
MO000162702	phlpp2(h)	PHLPP2	PH domain and leucine rich repeat protein phosphatase 2	1	182	0.38
MO000020073	Ubc5A(h)	UBE2D1	ubiquitin conjugating enzyme E2 D1	1	186	0.42
MO000022059	LynB(h)	LYN	LYN proto-oncogene, Src family tyrosine kinase	1	191	0.46

Table 10. Master regulators that may govern the regulation of **down-regulated** genes in Myc\_induce vs. Control. **Total rank** is the sum of the ranks of the master molecules sorted by keynode score, CMA score, transcriptomics and proteomics data. **See full table**  $\rightarrow$ 

ID	Master molecule name	Gene symbol	Gene description	Contained in proteome set		logFC (transcriptome)
MO000032768	ILK(h)	ILK	integrin linked kinase	1	133	-0.75
MO000033313	PKACA(h)	PRKACA	protein kinase cAMP- activated catalytic subunit alpha	1	218	-1.18
MO000021736	Cdk2(h)	CDK2	cyclin dependent kinase 2	1	224	-0.51
MO000021208	Caspase-6(h)	CASP6	caspase 6	1	233	-0.73
MO000017291	integrins	ITGA1, ITGA2B, ITGA3, ITGA4, ITGA5, ITGA6, ITGA8, ITGA9, ITGAL, ITGAV, ITGB1, ITGB2, ITGB3, ITGB4, I	integrin subunit alpha 1, integrin subunit alpha 2b, integrin subunit alpha 3, integrin subunit alph	1	236	-1.41
MO000099197	ILK-isoform1(h)	ILK	integrin linked kinase	1	240	-0.75
MO000079043	PML-4(h)	PML	promyelocytic leukemia	1	252	-0.51
MO000032135	proCaspase-6(h)	CASP6	caspase 6	1	306	-0.73
MO000124674	EPHB2(h)	EPHB2	EPH receptor B2	1	335	-0.91
MO000118076	EGF:ErbB1{pY}:ErbB2{pY}:Src	EGF, EGFR, ERBB2, SRC	SRC proto-oncogene, non- receptor tyrosine kinase, epidermal growth factor, epidermal growth factor r	1	336	-0.93

The intracellular regulatory pathways controlled by the above-mentioned master regulators are depicted in Figures 15 and 16. These diagrams display the connections between identified transcription factors, which play important roles in the regulation of differentially expressed genes, and selected master regulators, which are responsible for the regulation of these TFs.

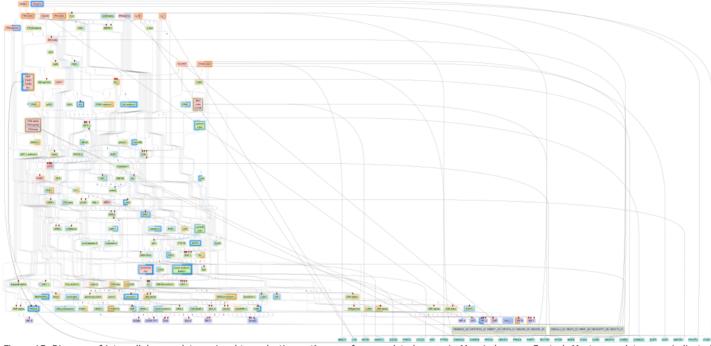


Figure 15. Diagram of intracellular regulatory signal transduction pathways of up-regulated genes in Myc\_induce vs. Control. Master regulators are indicated by red rectangles, transcription factors are blue rectangles, and green rectangles are intermediate molecules, which have been added to the network during the search for master regulators from selected TFs. Orange and blue frames highlight molecules that are encoded by up- and downregulated genes, resp. The left half of a highlighting frame corresponds to transcriptomic data, the right one to proteomic data. See full diagram  $\rightarrow$ 

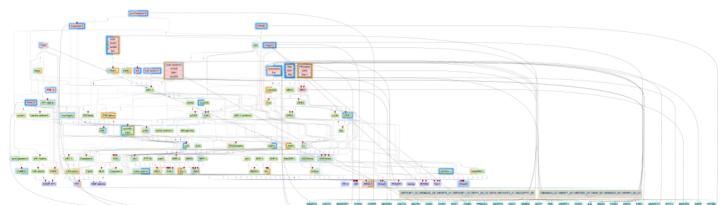


Figure 16. Diagram of intracellular regulatory signal transduction pathways of down-regulated genes in Myc\_induce vs. Control. Master regulators are indicated by red rectangles, transcription factors are blue rectangles, and green rectangles are intermediate molecules, which have been added to the network during the search for master regulators from selected TFs. Orange and blue frames highlight molecules that are encoded by up- and downregulated genes, resp. The left half of a highlighting frame corresponds to transcriptomic data, the right one to proteomic data. See full diagram  $\rightarrow$ 

## 4. Identification of potential drugs

In the last step of the analysis we strived to identify known drugs as well as new potentially active chemical compounds that are potentially suitable for inhibition (or activation) of the identified molecular targets in the context of specified human disease.

First, we identify known drugs using information from HumanPSD<sup>™</sup> database [5] about their targets and about clinical trials where the drugs have been tested for the treatment of various human diseases. Table 11 shows the resulting list of druggable master regulators that represent the predicted drug targets of the studied pathology. Table 12 lists chemical compounds and known drugs (from the HumanPSD<sup>™</sup> database) potentially acting on corresponding master regulators.

Table 11. Known drug targets for known drugs revealed in this study. The column **Druggability score** contains the number of drugs that are potentially suitable for inhibition (or activation) of the target. **Total rank** is the sum of the ranks of the master molecules sorted by keynode score, CMA score, transcriptomics and proteomics data. **See full table**  $\rightarrow$ 

ID	Gene symbol	Gene description	Druggability score	Contained in proteome set	Total rank	logFC (transcriptome)
ENSG00000134058	CDK7	cyclin dependent kinase 7	2	1	152	0.63
ENSG00000254087	LYN	LYN proto-oncogene, Src family tyrosine kinase	4	1	192	0.46
ENSG00000104365	ІКВКВ	inhibitor of nuclear factor kappa B kinase subunit beta	7	1	211	0.23
ENSG00000104695	PPP2CB	protein phosphatase 2 catalytic subunit beta	1	1	231	0.21
ENSG00000163932	PRKCD	protein kinase C delta	2	1	231	0.86
ENSG0000065613	SLK	STE20 like kinase	2	1	243	0.4
ENSG00000115232	ITGA4	integrin subunit alpha 4	8	1	247	0.51
ENSG00000169032	MAP2K1	mitogen-activated protein kinase kinase 1	10	1	268	0.22
ENSG00000153208	MERTK	MER proto-oncogene, tyrosine kinase	1	0	278	0.95
ENSG0000070770	CSNK2A2	casein kinase 2 alpha 2	1	1	298	0.47

Table 12. The list of drugs (from Human PSD) approved or used in clinical trials for the application in neoplasm metastasis and osteosarcoma and acting on master regulators revealed in our study. The column **Target activity score** contains the value of numeric function that depends on ranks of all targets that were found for the drug. The column **Disease activity score** contains the weighted sum of user selected diseases where the drug is known to be applied. We use sum of clinical trials phases as the weight of the disease. **Drug rank** column contains total rank of given drug among all found. See Methods section for details. **See full table**  $\rightarrow$ 

ID	Name	Target names	Target activity score	NA	Phase 1	Phase 2	Phase 3	Phase 4	[ a s
DB06616	Bosutinib	SRC, MAP2K1, LYN	0.96	Breast Neoplasms, Leukemia, Lymphoid, Leukemia, Myelogenous, Chronic, BCR-ABL Positive, Leukemia, Myeloid, Neoplasms, Precursor Cell Lymphoblastic Leukemia- Lymphoma	Acute Kidney Injury, Breast Neoplasms, Carcinoma, Non- Small-Cell Lung, Cholangiocarcinoma, Cognitive Dysfunction, Colorectal Neoplasms, Dementia	Neoplasm Metastasis, Brain Abscess, Breast Neoplasms, Cholangiocarcinoma, Colorectal Neoplasms, Cysts, Glioblastoma	Leukemia, Leukemia, Myelogenous, Chronic, BCR- ABL Positive, Leukemia, Myeloid	Leukemia, Myeloid	2
DB01269	Panitumumab	EGFR	0.39	Adenocarcinoma, Carcinoma, Squamous Cell, Carcinoma, Transitional Cell, Colorectal Neoplasms, Histology, Neoplasms	Adenocarcinoma, Carcinoma, Non- Small-Cell Lung, Carcinoma, Small Cell, Carcinoma, Squamous Cell, Colonic Neoplasms, Colorectal Neoplasms, Dermoid Cyst	Neoplasm Metastasis, Adenocarcinoma, Adenoma, Adenoma, Pleomorphic, Biliary Tract Neoplasms, Breast Neoplasms, Carcinoid Tumor	Colorectal Neoplasms, Neoplasms, Squamous Cell, Noma, Rectal Neoplasms, Stomach Neoplasms	Neoplasms	2
DB00317	Gefitinib	EGFR	0.39	Adenocarcinoma, Bronchopulmonary Dysplasia, Carcinoma, Adenoid Cystic, Carcinoma, Basal Cell, Carcinoma, Medullary, Carcinoma, Mucoepidermoid	Osteosarcoma, Astrocytoma, Breast Neoplasms, Carcinoma, Non- Small-Cell Lung, Carcinoma, Renal Cell, Carcinoma, Small Cell, Carcinoma, Squamous Cell	Adenocarcinoma, Adenocarcinoma, Mucinous, Adenoma, Astrocytoma, Brain Neoplasms, Breast Neoplasms, Breast Neoplasms, Male	Adenocarcinoma, Breast Neoplasms, Carcinoma, Carcinoma, Non- Small-Cell Lung, Carcinoma, Small Cell, Carcinoma, Squamous Cell, Colorectal Neoplasms	Carcinoma, Non-Small-Cell Lung, Carcinoma, Small Cell, Lung Neoplasms, Neoplasms, Noma, Small Cell Lung Carcinoma	1
DB08881	Vemurafenib	BRAF	0.32	Melanoma, Neoplasms, Noma, Thyroid Neoplasms	Colorectal Neoplasms, Glioma, Lymphoma, Melanoma, Neoplasms, Noma, Rectal Neoplasms	Osteosarcoma, Anger, Brain Abscess, Carcinoma, Non- Small-Cell Lung, Carcinoma, Small Cell, Colorectal Neoplasms, Craniopharyngioma	Melanoma, Noma	Melanoma, Neoplasms, Noma	2
DB06151	Acetylcysteine	ІКВКВ, СНИК	0.35	Acute Kidney Injury, Acute Lung Injury, Albuminuria, Aneurysm, Apnea, Bipolar Disorder, Bites and Stings	Osteosarcoma, Albinism, Albinism, Oculocutaneous, Alcoholism, Altitude Sickness, Amphetamine- Related Disorders, Anemia	Acute Kidney Injury, Acute Lung Injury, Adrenoleukodystrophy, Albinism, Albinism, Oculocutaneous, Alcohol Drinking, Alcoholism	Acute Kidney Injury, Anemia, Sickle Cell, Atrial Fibrillation, Atrophy, Bacteremia, Brain Death, Brain Diseases	Acute Kidney Injury, Alcoholism, Anemia, Atherosclerosis, Atrophy, Bipolar Disorder, Bronchiectasis	1

Table 13. The list of drugs (from HumanPSD) known to be acting on master regulators revealed in our study that can be proposed as a drug repurposing initiative for the treatment of neoplasm metastasis and osteosarcoma. **Target activity score** column contains value of numeric function that depends on ranks of all targets that were found for the drug. **Drug rank** column contains total rank of given drug among all found. See <u>Methods</u> section for details.

ID	Name	Target names	Target activity score	NA	Phase 1	Phase 2	Phase 3	Phase 4	Drug rank
DB05013	Ingenol Mebutate	PRKCD, PRKCA	0.87	Keratosis, Keratosis, Actinic	Keratosis, Keratosis, Actinic, Warts	Carcinoma, Basal Cell, Keratosis, Keratosis, Actinic, Keratosis, Seborrheic, Noma, Sunburn	Keratosis, Keratosis, Actinic	Keratosis, Keratosis, Actinic	24
DB00163	Vitamin E	PPP2CB, PRKCA, PPP2CA	0.61	Apnea, Fatty Liver, Fatty Liver, Alcoholic, Fragile X Syndrome, HIV Infections, Hepatitis, Infection	Alzheimer Disease, Anemia, Anemia, Iron- Deficiency, Arthritis, Arthritis, Rheumatoid, Asthma, Brain Abscess	Acute Kidney Injury, Adrenoleukodystrophy, Anemia, Arsenic Poisoning, Arteriosclerosis, Atherosclerosis, Brain Abscess	Alzheimer Disease, Angina, Unstable, Arterial Occlusive Diseases, Arteriosclerosis, Burns, Cardiovascular Diseases, Cataract	Angina Pectoris, Variant, Asphyxia, Cicatrix, Cicatrix, Hypertrophic, Diabetes Mellitus, Dyslipidemias, Epilepsy	28
DB09033	Vedolizumab	ITGA4	0.48	Cholangitis, Cholangitis, Sclerosing, Colitis, Colitis, Ulcerative, Crohn Disease, Graft vs Host Disease, Inflammatory Bowel Diseases	Colitis, Colitis, Ulcerative, Crohn Disease, Inflammatory Bowel Diseases, Melanoma, Noma, Ulcer	Celiac Disease, Colitis, Colitis, Ulcerative, Crohn Disease, Ulcer	Cholangitis, Cholangitis, Sclerosing, Colitis, Colitis, Ulcerative, Crohn Disease, Inflammatory Bowel Diseases, Ulcer	Colitis, Colitis, Ulcerative, Crohn Disease, Ulcer	32
DB08911	Trametinib	MAP2K1	0.44	Adenocarcinoma, Biliary Tract Neoplasms, Brain Abscess, Carcinoma, Non- Small-Cell Lung, Carcinoma, Small Cell, Gastrointestinal Stromal Tumors, Germinoma	Adenocarcinoma, Adenocarcinoma, Clear Cell, Behavior, Brain Abscess, Breast Neoplasms, Carcinoma, Carcinoma, Non- Small-Cell Lung	Adenocarcinoma, Astrocytoma, Bile Duct Neoplasms, Brain Abscess, Breast Neoplasms, Carcinoma, Hepatocellular, Carcinoma, Non- Small-Cell Lung	Adenocarcinoma, Melanoma, Noma	Carcinoma, Small Cell, Glioma, Lung Neoplasms, Melanoma, Neoplasms, Noma, Small Cell Lung Carcinoma	39
DB00244	Mesalazine	ІКВКВ, СНИК	0.44	Abdominal Pain, Character, Colitis, Colitis, Collagenous, Colitis, Ulcerative, Crohn Disease, Diarrhea		Adenoma, Colitis, Colitis, Ulcerative, Colorectal Neoplasms, Crohn Disease, Digestive System Diseases, Gastrointestinal Diseases	Colitis, Colitis, Collagenous, Colitis, Ulcerative, Crohn Disease, Digestive System Diseases, Diverticulitis, Gastrointestinal Diseases	Colitis, Colitis, Ulcerative, Diarrhea, Diverticulum, Irritable Bowel Syndrome, Ulcer	47

Next, new potential small molecular ligands were predicted for the revealed targets and a general druggability check was run using a precomputed database of spectra of biological activities of chemical compounds from a library of 13040 most pharmaceutically active known compounds. The spectra of biological activities has been computed using the program PASS [11-13] on the basis of a (Q)SAR approach. Table 14 shows the resulting list of druggable master regulators, which represent the predicted drug targets of the studied pathology. Table 15 lists chemical compounds and known drugs potentially acting on the corresponding master regulators.

Table 14. Extended list of drug targets revealed in this study (targets that are predicted by PASS program potentially targeted by an extended list of known drugs and pharmaceutically active chemical compounds). The column **Druggability score** contains a numeric value which indicates how suitable this target is to be inhibited (or activated) by a drug. See Methods section for details. **See full table**  $\rightarrow$ 

ID	Name	Gene symbol	Gene description	Druggability score	Contained in proteome set	Total rank	logFC (transcriptome)
ENSG0000020426	MNAT1	MNAT1	MNAT1, CDK activating kinase assembly factor	17.43	1	152	0.63
ENSG00000134058	CDK7	CDK7	cyclin dependent kinase 7	11.42	1	152	0.63
ENSG00000134480	CCNH	CCNH	cyclin H	9.81	1	152	0.63
ENSG00000104312	RIPK2	RIPK2	receptor interacting serine/threonine kinase 2	1.41	1	192	0.42
ENSG00000254087	LYN	LYN	LYN proto-oncogene, Src family tyrosine kinase	7.71	1	192	0.46
ENSG00000171132	PRKCE	PRKCE	protein kinase C epsilon	62.09	1	194	0.5
ENSG00000104365	IKBKB	ІКВКВ	inhibitor of nuclear factor kappa B kinase subunit beta	1.6	1	211	0.23
ENSG00000145335	SNCA	SNCA	synuclein alpha	1.74	1	222	8.69E-2
ENSG00000163932	PRKCD	PRKCD	protein kinase C delta	62.73	1	231	0.86
ENSG00000100393	EP300	EP300	E1A binding protein p300	15.7	1	240	0.26

Table 15. The chemical compounds and known drugs identified by the PASS program as potentially active for the treatment of neoplasm metastasis and osteosarcoma and acting on master regulators revealed in our study. **Disease activity score** column contains maximal value of probability to be active for all activities corresponding to the selected diseases for the given compound. **Target activity score** column contains value of numeric function which depends on all activity-mechanisms correspondent to the drug. **Drug rank** column contains total rank of given drug among all found. See Methods section for details.

See full table $\rightarrow$					
Name	Structure	Target names	Target activity score	Disease activity score	Drug rank
Clomocycline		CSNK1G1, SIRT1	0.13	0.9	201
1-Deoxy-1-Methoxycarbamido-Beta-D- Glucopyranose		CARM1, ITGA4, ITGA6, IGF1R	3.67E-3	0.83	504

Table 16. The chemical compounds and known drugs identified by the PASS program as potentially acting on master regulators revealed in our study. Based on the revealed mechanism of action these compounds can be proposed for the treatment of neoplasm metastasis and osteosarcoma in the current pathological case. **Disease activity score** column contains maximal value of probability to be active for all activities corresponding to the selected diseases for the given compound or 0 if no diseases were selected (in this case column will be hidden). **Target activity score** column contains value of numeric function which depends on all activity-mechanisms correspondent to the drug. **Drug rank** column contains total rank of given drug among all found. See Methods section for details.

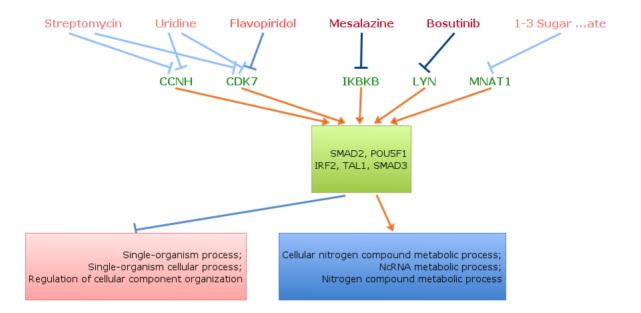
Name	Structure	Target names	Target activity score	Disease activity score	Drug rank
2'- Deoxymaltose		PRKCE, PRKCD, PRKCI, PRKCA	0.38	0.74	55
SU9516		MTOR, PRKCE, PRKCD, PRKCI, PRKCA	0.92	0.5	69
Cholic Acid	рики страна и стран	EGFR, GRK2, PRKAA2, MERTK, ROCK1, PKMYT1, IGF1R	0.38	0.62	71
Xanthophyll		PRKCE, PRKCD, PRKCI, PRKCA	0.46	0.55	80
Streptomycin	$ \begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & $	CDK6, CCNH, CCND3, PRKCE, PRKCD, PRKCI, PRKCA	0.75	0.49	80

As a result of the drug search we came up with two lists of chemical compounds potentially applicable to the targets of our interest. The first list is based on drugs that are known as ligands for the revealed targets in the context of the diseases in our focus as well as in other disease conditions. The second list of identified compounds is based on the prediction of their potential biological activities, which was done using the program PASS. Such computational predictions should be taken as mere suggestions and should be used with care in further experiments.

## 5. Conclusion

We applied the software package "Genome Enhancer" to a multi-omics data set that contains *transcriptomics and proteomics* data. The study is done in the context of *neoplasm metastasis and osteosarcoma*. The data were pre-processed, statistically analyzed and differentially expressed genes were identified. Also checked was the enrichment of GO or disease categories among the studied gene sets.

We propose the following schema of how the selected drugs may interfere with the identified target molecules and pathogenic processes discovered by the study reported here.



## 6. Methods

#### Databases used in the study

Transcription factor binding sites in promoters and enhancers of differentially expressed genes were analyzed using known DNA-binding motifs described in the TRANSFAC® library, release 2020.1 (geneXplain GmbH, Wolfenbüttel, Germany) (http://genexplain.com/transfac). The master regulator search uses the TRANSPATH® database (BIOBASE), release 2020.1 (geneXplain GmbH, Wolfenbüttel, Germany) (http://genexplain.com/transpath). A comprehensive signal transduction network of human cells is built by the software on the basis of reactions annotated in TRANSPATH®.

The information about drugs corresponding to identified drug targets and clinical trials references were extracted from HumanPSD<sup>™</sup> database, release 2020.1 (http://genexplain.com/humanpsd).

The Ensembl database release Human88.38 (hg38) (http://www.ensembl.org) was used for gene IDs representation and Gene Ontology (GO) (http://geneontology.org) was used for functional classification of the studied gene set.

#### Methods for the analysis of enriched transcription factor binding sites and composite modules

Transcription factor binding sites in promoters and enhancers of differentially expressed genes were analyzed using known DNA-binding motifs. The motifs are specified using position weight matrices (PWMs) that give weights to each nucleotide in each position of the DNA binding motif for a transcription factor or a group of them.

We search for transcription factor binding sites (TFBS) that are enriched in the promoters and enhancers under study as compared to a background sequence set such as promoters of genes that were not differentially regulated under the condition of the experiment. We denote study and background sets briefly as Yes and No sets. In the current work we used a workflow considering promoter sequences of a standard length of 1100 bp (-1000 to +100). The error rate in this part of the pipeline is controlled by estimating the adjusted p-value (using the Benjamini-Hochberg procedure) in comparison to the TFBS frequency found in randomly selected regions of the human genome (adj.p-value < 0.01).

We have applied the CMA algorithm (Composite Module Analyst) for searching composite modules [7] in the promoters and enhancers of the Yes and No sets. We searched for a composite module consisting of a cluster of 10 TFs in a sliding window of 200-300 bp that statistically significantly separates sequences in the Yes and No sets (minimizing Wilcoxon p-value).

#### Methods for finding master regulators in networks

We searched for master regulator molecules in signal transduction pathways upstream of the identified transcription factors. The master regulator search uses a comprehensive signal transduction network of human cells. The main algorithm of the master regulator search has been described earlier [3,4]. The goal of the algorithm is to find nodes in the global signal transduction network that may potentially regulate the activity of a set of transcription factors found at the previous step of the analysis. Such nodes are considered as most promising drug targets, since any influence on such a node may switch the transcriptional programs of hundreds of genes that are regulated by the respective TFs. In our analysis, we have run the algorithm with a maximum radius of 12 steps upstream of each TF in the input set. The error rate of this algorithm is controlled by applying it 10000 times to randomly generated sets of input transcription factors of the same set-size. Z-score and FDR value of ranks are calculated then for each potential master regulator node on the basis of such random runs (see detailed description in [9]). We control the error rate by the FDR threshold 0.05.

#### Methods for analysis of pharmaceutical compounds

We seek for the optimal combination of molecular targets (key elements of the regulatory network of the cell) that potentially interact with pharmaceutical compounds from a library of known drugs and biologically active chemical compounds, using information about known drugs from HumanPSD<sup>™</sup> and predicting potential drugs using PASS program.

Method for analysis of known pharmaceutical compounds

We selected compounds from HumanPSD<sup>™</sup> database that have at least one target. Next, we sort compounds using "*Drug rank*" that is sum of three other ranks:

- 1. ranking by "Target activity score" (*T*-score<sub>PSD</sub>),
- 2. ranking by "Disease activity score" (*D*-score<sub>PSD</sub>),
- 3. ranking by clinical trials phase.

To calculate clinical trials phase for the given compound we select the maximum phase of all diseases that are known to have clinical trials with this compound. "Target activity score" (*T*-score<sub>PSD</sub>) is calculated as follows:

$$T\text{-}score_{\scriptscriptstyle PSD} = -\frac{|T|}{|T| + w(|AT| - |T|))} \sum_{t \in T} \log_{10} \left(\frac{rank(t)}{1 + maxRank(T)}\right)$$

where *T* is set of all targets related to the compound intersected with input list, |T| is number of elements in *T*, *AT* and |AT| are set set of all targets related to the compound and number of elements in it, *w* is weight multiplier, *rank*(*t*) is rank of given target, *maxRank*(*T*) equals max(rank(t)) for all targets *t* in *T*.

We use following formula to calculate "Disease activity score" ( D-score<sub>PSD</sub>):

$$D\text{-}score_{\scriptscriptstyle PSD} = \begin{cases} \sum\limits_{d \in D} \sum\limits_{p \in P} phase(d, p) \\ 0, \ D = \varnothing \end{cases}$$

where *D* is the set of selected diseases, and if *D* is empty set, D-score<sub>*PSD*</sub>=0. *P* is a set of all known phases for each disease, *phase*(*p*,*d*) equals to the phase number if there are known clinical trials for the selected disease on this phase and zero otherwise.

#### Method for prediction of pharmaceutical compounds

In this study, the focus was put on compounds with high pharmacological efficiency and low toxicity. For this purpose, comprehensive library of chemical compounds and drugs was subjected to a SAR/QSAR analysis. This library contains 13040 compounds along with their pre-calculated potential pharmacological activities of those substances, their possible side and toxic effects, as well as the possible mechanisms of action. All biological activities are expressed as probability values for a substance to exert this activity (*Pa*).

We selected compounds that satisfied the following conditions:

- 1. Toxicity below a chosen toxicity threshold (defines as Pa, probability to be active as toxic substance).
- 2. For all predicted pharmacological effects that correspond to a set of user selected disease(s) *Pa* is greater than a chosen effect threshold.
- 3. There are at least 2 targets (corresponding to the predicted activity-mechanisms) with predicted *Pa* greater than a chosen target threshold.

The maximum *Pa* value for all toxicities corresponding to the given compound is selected as the "Toxicity score". The maximum *Pa* value for all activities corresponding to the selected diseases for the given compound is used as the "Disease activity score". "Target activity score" (T-score) is calculated as follows:

$$T\text{-}score(s) = \frac{|T|}{|T| + w(|AT| - |T|))} \sum_{m \in \mathcal{M}(s)} \left( pa(m) \sum_{g \in G(m)} IAP(g)optWeight(g) \right)$$

where M(s) is the set of activity-mechanisms for the given structure (which passed the chosen threshold for activity-mechanisms Pa); G(m) is the set of targets (converted to genes) that corresponds to the given activity-mechanism (m) for the given compound; pa(m) is the probability to be active of the activity-mechanism (m), IAP(g) is the invariant accuracy of prediction for gene from G(m); optWeight(g) is the additional weight multiplier for gene. T is set of all targets related to the compound intersected with input list, |T| is number of elements in T, AT and |AT| are set set of all targets related to the compound and number of elements in it, w is weight multiplier.

"Druggability score" (D-score) is calculated as follows: D-score $(g) = IAP(g) \sum_{a} \sum_{b} pa(m),$ 

$$pre(g) = IAP(g) \sum_{s \in S(g)} \sum_{m \in M(s,g)} pa(m)$$

where S(g) is the set of structures for which target list contains given target, M(s,g) is the set of activity-mechanisms (for the given structure) that corresponds to the given gene, pa(m) is the probability to be active of the activity-mechanism (m), IAP(g) is the invariant accuracy of prediction for the given gene.

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#### Thank you for using the Genome Enhancer!

In case of any questions please contact us at <a href="mailto:support@genexplain.com">support@genexplain.com</a>

#### Supplementary material

- 1. Supplementary table 1 Up-regulated genes
- 2. Supplementary table 2 Down-regulated genes
- 3. Supplementary table 3 Detailed report. Composite modules and master-regulators (up-regulated genes in Myc\_induce vs. Control).
- 4. Supplementary table 4 Detailed report. Composite modules and master-regulators (down-regulated genes in Myc\_induce vs. Control).
- 5. Supplementary table 5 Detailed report. Pharmaceutical compounds and drug targets.

#### Disclaimer

Decisions regarding care and treatment of patients should be fully made by attending doctors. The predicted chemical compounds listed in the report are given only for doctor's consideration and they cannot be treated as prescribed medication. It is the physician's responsibility to independently decide whether any, none or all of the predicted compounds can be used solely or in combination for patient treatment purposes, taking into account all applicable information regarding FDA prescribing recommendations for any therapeutic and the patient's condition, including, but not limited to, the patient's and family's medical history, physical examinations, information from various diagnostic tests, and patient preferences in accordance with the current standard of care. Whether or not a particular patient will benefit from a selected therapy is based on many factors and can vary significantly.

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